Synthesis of New 1,2,4-Triazole Derivatives and Investigation of Their Matrix Metalloproteinase-9 (MMP-9) Inhibition Properties

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ABSTRACT

Nine new 2-[[5-((4-acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(aryl/heteroaryl)acetamide (5a-5i) derivatives were synthesized and tested for their matrix metalloproteinase-9 (MMP-9) inhibition potency which is associated with antiproliferative activity. None of the compounds exhibited MMP-9 inhibition activity close to standard drug however, two compounds 5e bearing 6-methylbenzothiazole and 5g bearing 6-ethoxybenzothiazole moieties showed the highest MMP-9 inhibition which were determined over than 50% percentage at 100 μg/mL concentration. Some physicochemical properties of all final compounds were calculated via SwissADME and the results were modest to be able to an oral drug. Molecular docking studies were realized with MMP-9 enzyme (PDB ID: 5I12) and 5d, 5e, and 5g compounds for comparing the active and non-active/low effective structures.

Keywords: 1,2,4-Triazole, matrix metalloproteinase-9 (MMP-9), antiproliferative activity, molecular docking

INTRODUCTION

Cancer is a common and tough disease which its many types still have no definite and radical treatment. In this regard, the disease has been tried to be controlled by understanding the damages it causes and focusing on targeted path-
ways related to the emergence or metastasis of the disease.\textsuperscript{1,2} Target therapy mostly includes a specific site, such as tumour vasculature or tumoral intracellular organelles which provides protection from the side effects of treatment and leaving the surrounding tissue unaffected. Additionally, gene therapy and expression of genes triggering apoptosis with(out) some enzymes and wild type tumour suppressors or the targeted silencing mediated by siRNAs are the other ways of targeted chemotherapy.\textsuperscript{3,4} Matrix metalloproteinase enzymes MMPs are zinc- and calcium-dependent endopeptidases involved in physiological and pathological processes such as embryo implantation, wound healing, tissue remodeling and formation of some malfunctions like cancer, arthritis, genetic disorders, chronic kidney, aging and cardiovascular diseases.

The MMP family consists of 25 secreted and membrane-bound members, among them MMP-2 (gelatinase A) and MMP-9 (gelatinase B) serve in inflammatory response and angiogenesis. It has been determined that gelatinases show tumorigenic activity by the way of degradation of the extracellular matrix (ECM) and non-ECM functions that activates pro-TNF-\(\alpha\) and TGF-\(\beta\) (transforming growth factor beta) molecules in tumor development stages, such as tumor cell growth, migration, invasion, and metastasis. The regulatory role of MMPs in cancer progression is a current strategy for treatment cancer disease.\textsuperscript{5-7}

Nitrogen containing heterocycles are valuable drug designing skeletons, among these 1,2,4 triazoles carrying three nitrogen atoms have considerable interest due to their synthetic easibility, electron-rich chemical properties and wide spectrum of biological applications such as antioxidant, anticonvulsant, anti-inflammatory, antimicrobial and antiviral activities. 1,2,4-triazole nucleus is also placed in drug candidates including \(H_1/H_2\) histamine receptor blockers, cholinesterase active agents, central nervous system (CNS) stimulants, antianxiety and sedatives along with a lot of clinically used drugs eg. alprazolam (tranquilizer), benatradin (diuretic), trapidil (hypotensive), trazodon (antidepressant, anxiolytic), ribavirin (antiviral), fluconazole, itraconazole, terconazole (antifungal), anastrozole, letrozole and vorozole (antineoplastics).\textsuperscript{8-12} The antiproliferative activity of 1,2,4-triazole derivative has been reported to associated with cell cycle arrest in the G\(_2\)/M phase via binding property to variable receptors and enzymes due to the ring’s polar nature. Additionally, other triazole including molecules have been determined to exhibit their anticancer potency via tubulin polymerization inhibition and modulation, mitochondrial depolarization, and induction of apoptosis through activation of caspase-3, inhibition of kinase, aromatase and steroid sulfatase, methionine aminopeptidase inhibitors and tankyrase enzymes.\textsuperscript{13,14}
Based on the reported data and as an expansion work of our previous study,\textsuperscript{15} nine 1,2,4-triazole derivatives were synthesized, inhibition of MMP-9 enzyme was investigated and structure-activity relationships of the molecules was discussed.

**METHODOLOGY**

**Chemistry**

All chemicals used in the syntheses were purchased either from Merck Chemicals (Merck KGaA, Darmstadt, Germany) or Sigma-Aldrich Chemicals (Sigma-Aldrich Corp., St. Louis, MO, USA). The reactions and the purity of the compounds were observed by thin-layer chromatography (TLC) on silica gel 60 F254 aluminum sheets obtained from Merck (Darmstadt, Germany). Melting points of the synthesized compounds were recorded by the MP90 digital melting point apparatus (Mettler Toledo, Ohio, USA) and were presented as uncorrected. \textsuperscript{1}H NMR and \textsuperscript{13}C NMR spectra were recorded by a Bruker 300 MHz and 75 MHz digital FT-NMR spectrometer (Bruker Bioscience, Billerica, MA, USA) in DMSO-\textit{d}_6, respectively. In the NMR spectra, splitting patterns were designated as follows: s: singlet; d: doublet; t: triplet; m: multiplet. Coupling constants (\(J\)) were reported as Hertz. High resolution mass spectrometric (HRMS) studies were performed using an LC/MS-IT-TOF system (Shimadzu, Kyoto, Japan). Elemental analyses were performed on a Leco 932 CHNS analyzer (Leco, Michigan, USA).

**Ethyl 2-(4-acetamidophenoxy)acetate (1)**

\(N-(4\text{-Hydroxyphenyl})\text{acetamide (0.033 mol, 5g)}, \text{ethyl chloroacetate (0.04 mol, 4.88 g)}\) and potassium carbonate were refluxed in acetone for 6h. After checking the reaction with TLC, the solvent was evaporated, and the material was filtered and washed with water. The raw product was crystallised from ethanol.

**N-[4-(2-Hydrazinyl-2-oxoethoxy)phenyl]acetamide (2)**

Ethyl 2-(4-acetamidophenoxy)acetate (1, 1 eq.) and hydrazine monohydrate (1.2 eq) were stirred in ethanol at room temperature overnight. When the reaction was over, the mixture was separated from the solvent. The precipitated material was filtered and crystallised from ethanol.

**N-(4-(2-Oxo-2-(2-(phenylcarbamothioyl)hydrazinyl)ethoxy)phenyl)acetamide (3)**

\(N-(4\text{-Hydrazinyl-2-oxoethoxy)phenyl)acetamide (2)}\) and phenylisothiocyanate were refluxed in EtOH for 3h. After TLC, the solvent was evaporated, and the solid was collected.
N-[4-((5-Mercapto-4-phenyl-4H-1,2,4-triazol-3-yl)methoxy)phenyl]acetamide (4)

N-(4-(2-oxo-2-(2-phenylcarbamothioyl)hydrazinyl)ethoxy)phenyl)acetamide (3) was refluxed with 2N potassium hydroxide prepared in ethanol for 2h. After TLC, the pH was adjusted to 7 in a cold environment to precipitate the material.

General procedure for the synthesis N-substituted-2-[[5-((4-acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]acetamide derivatives (5a-5i)

N-(4-((5-mercapto-4-phenyl-4H-1,2,4-triazol-3-yl)methoxy)phenyl)acetamide (4) (0.0015 mol, 0.5 g) and equivalent 2-chloro-N-substituted acetamide derivatives were reacted in the presence of potassium carbonate in acetone. After 4-5 h, it was finished, the solvent was evaporated and treated with water, then it was crystallised from ethanol to yield the final product.

2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(6-nitrobenzothiazol-2-yl)acetamide (5a)

m. p. 240°C, ¹H-NMR (300 MHz, DMSO-đ, ppm) δ 1.98 (s, 3H, CH₃), 4.33 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.77 (d, J=8.98 Hz, 2H, Ar-H), 7.40 (d, J=8.98 Hz, 2H, Ar-H), 7.48-7.64 (m, 5H, Ar-H), 7.54 (d, J=8.95 Hz, 1H, Ar-H), 8.27 (dd, J₁=2.43 Hz, J₂= 8.94 Hz, 1H, Ar-H), 9.03 (m, 1H, Ar-H), 9.79 (brs, 1H, NH). ¹³C-NMR (75 MHz, DMSO-đ, ppm) δ 24.26 (C-CH₃), 36.54 (S-CH₂), 60.55 (O-CH₂), 115.45, 119.40, 120.76, 121.11, 122.25, 126.66, 127.43, 130.29, 130.64, 133.86, 143.21, 151.76, 152.38, 153.46, 154.22, 164.81, 168.27 (acetamide C=O), 168.85 (C=O). For C₂₆H₂₁N₇O₅S₂ calculated: 54.25 % C, 3.68 % H, 17.03 % N; found: 54.27 % C, 3.69 % H, 17.05 % N. HRMS [M-H]⁻: calculated: 574.0973, experimental: 574.1010.

2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(benzothiazol-2-yl)acetamide (5b)

m. p. 270°C, ¹H-NMR (300 MHz, DMSO-đ, ppm) δ 1.99 (s, 3H, CH₃), 4.31 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.77 (d, J=9.02 Hz, 2H, Ar-H), 7.30-7.60 (m, 9H, Ar-H), 7.77 (d, J=8.03 Hz, 1H, Ar-H), 7.80 (d, J=7.86 Hz, 1H, Ar-H), 9.78 (brs, 1H, NH), 12.70 (brs, 1H, NH). ¹³C-NMR (75 MHz, DMSO-đ, ppm) δ 24.28 (C-CH₃), 36.08 (S-CH₂), 60.56 (O-CH₂), 115.45, 120.76, 120.89, 122.17, 127.43, 130.29, 130.64, 130.91, 133.00, 133.87, 151.66, 152.40, 153.47, 167.45 (acetamide C=O), 168.24 (C=O). For C₂₆H₂₂N₆O₃S₂ calculated: 58.85 % C, 4.18 % H, 15.84 % N; found: 58.82 % C, 4.19 % H, 15.85 % N. HRMS [M-H]⁻: calculated: 529.1122, experimental: 529.1153.
2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(6-fluorobenzothiazol-2-yl)acetamide (5c)

m. p. 276°C, ¹H-NMR (300 MHz, DMSO-d₆, ppm) δ 2.00 (s, 3H, CH₃), 4.31 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.77 (dd, J₁=1.97 Hz, J₂=7.08 Hz, 2H, Ar-H), 7.27-7.33 (m, 1H, Ar-H), 7.40 (dd, J₁=2.01 Hz, J₂=7.08 Hz, 2H, Ar-H), 7.50-7.58 (m, 5H, Ar-H), 7.78 (dd, J₁=4.84 Hz, J₂=8.88 Hz, 1H, Ar-H), 7.89 (dd, J₁=2.65 Hz, J₂=8.7 Hz, 1H, Ar-H), 9.79 (brs, 1H, NH), 12.73 (brs, 1H, NH).

¹³C-NMR (75 MHz, DMSO-d₆, ppm) δ 24.24 (CH₃), 36.02 (S-CH₂), 60.56 (O-CH₂), 108.53, 108.89, 114.63, 114.96, 115.45, 120.76, 122.32, 127.43, 130.29, 130.64, 131.00, 133.09, 133.23, 133.88, 145.69, 151.64, 152.42, 153.47, 157.57, 158.19, 160.75, 167.54 (acetamide C=O), 168.26 (C=O). For C₂₆H₂₁FN₆O₃S₂ calculated: 56.92 % C, 3.86 % H, 15.32 % N; found: 56.94 % C, 3.85 % H, 15.35 % N. HRMS [M-H]⁻: calculated: 547.1028, experimental: 547.1075.

2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(6-chlorobenzothiazol-2-yl)acetamide (5d)

m. p. 258°C, ¹H-NMR (300 MHz, DMSO-d₆, ppm) δ 1.98 (s, 3H, CH₃), 4.30 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.77 (d, J=9.03 Hz, 2H, Ar-H), 7.40 (d, J=9.01 Hz, 2H, Ar-H), 7.46 (dd, J₁=2.23 Hz, J₂=8.61 Hz, 1H, Ar-H), 7.50-7.58 (m, 5H, Ar-H), 7.76 (d, J=8.66 Hz, 1H, Ar-H), 8.13 (d, J=2.13 Hz, 1H, Ar-H), 9.79 (brs, 1H, NH), 12.80 (brs, 1H, NH).

¹³C-NMR (75 MHz, DMSO-d₆, ppm) δ 24.27 (C-CH₃), 36.06 (S-CH₂), 60.55 (O-CH₂), 115.45, 120.77, 121.97, 122.33, 127.00, 127.42, 128.16, 130.29, 130.65, 132.99, 133.62, 133.87, 147.88, 151.63, 152.41, 153.46, 159.14, 167.40 (acetamide C=O), 168.25 (C=O). For C₂₆H₂₁ClN₆O₃S₂ calculated: 55.27 % C, 3.75 % H, 14.87 % N; found: 55.28 % C, 3.72 % H, 14.86 % N. HRMS [M-H]⁻: calculated: 563.0732, experimental: 563.0768.

2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(6-methylbenzothiazol-2-yl)acetamide (5e)

m. p. 280°C, ¹H-NMR (300 MHz, DMSO-d₆, ppm) δ 1.98 (s, 3H, CH₃), 2.40 (s, 3H, CH₃), 4.30 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.78 (d, J=9.04 Hz, 2H, Ar-H), 7.26 (dd, J₁=1.25 Hz, J₂=7.30 Hz, 1H, Ar-H), 7.41 (d, J=9.10 Hz, 2H, Ar-H), 7.49-7.59 (m, 5H, Ar-H), 7.65 (d, J=8.28 Hz, 1H, Ar-H), 7.76 (s, 1H, Ar-H), 9.79 (brs, 1H, NH), 12.63 (brs, 1H, NH).

¹³C-NMR (75 MHz, DMSO-d₆, ppm) δ 24.27 (C-CH₃), 36.08 (S-CH₂), 60.55 (O-CH₂), 115.44, 120.76, 121.82, 127.43, 127.98, 130.28, 130.63, 132.05, 133.00, 133.60, 133.87, 146.02, 151.68, 152.40, 153.47, 157.39, 167.30 (acetamide C=O), 168.25 (C=O). For C₂₇H₂₄N₂O₃S₂ calculated: 59.54 % C, 4.44 % H, 15.43 % N; found: 59.50 % C, 4.42 % H, 15.45 % N. HRMS [M-H]⁻: calculated: 543.1279, experimental: 543.1326.
2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(6-methoxybenzothiazol-2-yl)acetamide (5f)

m. p. 256°C, 1H-NMR (300 MHz, DMSO-d₆, ppm) δ 1.98 (s, 3H, CH₃), 3.80 (s, 3H, O-CH₃), 4.29 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.77 (d, J=9.03 Hz, 2H, Ar-H), 7.04 (dd, J₁=2.58 Hz, J₂= 8.85 Hz, 1H, Ar-H), 7.40 (d, J= 8.98 Hz, 2H, Ar-H), 7.50-7.58 (m, 5H, Ar-H), 7.66 (d, J= 8.85 Hz, 1H, Ar-H), 9.79 (brs, 1H, NH), 12.57 (brs, 1H, NH). 13C-NMR (75 MHz, DMSO-d₆, ppm) δ 24.27 (C-CH₃), 36.04 (S-CH₂), 56.08 (O-CH₃), 60.56 (O-CH₂), 105.18, 115.45, 120.76, 121.72, 127.43, 127.98, 130.28, 130.64, 133.00, 133.24, 133.87, 143.04, 151.69, 152.39, 153.47, 156.21, 156.65, 167.13 (acetamide C=O), 168.25 (C=O). For C₂₇H₂₄N₆O₄S₂ calculated: 57.84 % C, 4.32 % H, 14.99 % N; found: 57.85 % C, 4.35 % H, 14.95 % N. HRMS [M-H]⁻: calculated: 559.1228, experimental: 559.1269.

2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(6-ethoxybenzothiazol-2-yl)acetamide (5g)

m. p. 226°C, 1H-NMR (300 MHz, DMSO-d₆, ppm) δ 1.34 (t, J=6.95 Hz, 3H, O-CH₂-CH₃), 1.97 (s, 3H, CH₃), 4.06 (q, J₁=6.93 Hz, J₂=13.85 Hz, 2H, O-CH₂-CH₃), 4.29 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.77 (d, J=8.97 Hz, 2H, Ar-H), 7.02 (dd, J₁=2.51 Hz, J₂= 8.82 Hz, 1H, Ar-H), 7.40 (d, J=8.96 Hz, 2H, Ar-H), 7.50-7.58 (m, 6H, Ar-H), 7.65 (d, J= 8.85 Hz, 1H, Ar-H), 9.79 (s, 1H, NH), 12.56 (brs, 1H, NH). 13C-NMR (75 MHz, DMSO-d₆, ppm) δ 15.17 (O-CH₂-CH₃), 24.25 (C-CH₃), 36.00 (O-CH₂), 64.04 (O-CH₂-CH₃), 105.81, 115.45, 115.82, 120.73, 127.43, 130.28, 130.64, 133.00, 133.24, 133.87, 142.93, 153.47, 155.80, 167.08 (acetamide C=O), 168.26 (C=O). For C₂₈H₂₆N₆O₄S₂ calculated: 58.52 % C, 4.56 % H, 14.62 % N; found: 58.53 % C, 4.55 % H, 14.65 % N. HRMS [M-H]⁻: calculated: 573.1384, experimental: 573.1436.

2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(4-nitrophenyl)acetamide (5h)

m. p. 250°C, 1H-NMR (300 MHz, DMSO-d₆, ppm) δ 1.98 (s, 3H, CH₃), 4.25 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.76-6.77 (m, 2H, Ar-H), 7.38-7.41 (m, 2H, Ar-H), 7.49-7.58 (m, 5H, Ar-H), 7.80-7.83 (m, 2H, Ar-H), 9.80 (brs, 1H, NH), 10.98 (brs, 1H, NH). 13C-NMR (75 MHz, DMSO-d₆, ppm) δ 24.27 (C-CH₃), 37.26 (S-CH₂), 60.51 (O-CH₂), 115.44, 120.74, 125.58, 130.29, 130.63, 133.00, 133.86, 142.81, 145.35, 152.30, 153.43, 167.00 (acetamide C=O), 168.26 (C=O). For C₂₅H₁₄N₂O₄S₂ calculated: 57.91 % C, 4.28 % H, 16.21 % N; found: 57.93 % C, 4.25 % H, 16.15 % N. HRMS [M-H]⁻: calculated: 517.1300, experimental: 517.1325.
2-[[5-((4-Acetamidophenoxy)methyl)-4-phenyl-4H-1,2,4-triazol-3-yl]thio]-N-(4-fluorophenyl)acetamide (5i)

m. p. 196°C, ¹H-NMR (300 MHz, DMSO-d₆, ppm) δ 1.99 (s, 3H, CH₃), 4.17 (s, 2H, S-CH₂), 5.03 (s, 2H, O-CH₂), 6.77 (d, J=8.99 Hz, 2H, Ar-H), 7.13-7.19 (m, 2H, Ar-H), 7.41 (d, J= 8.98 Hz, 2H, Ar-H), 7.48-7.51 (m, 2H, Ar-H), 7.56-7.58 (m, 5H, Ar-H), 9.80 (brs, 1H, NH), 10.42 (brs, 1H, NH). ¹³C-NMR (75 MHz, DMSO-d₆, ppm) δ 24.25 (C-CH₃), 37.16 (S-CH₂), 60.53 (O-CH₂), 115.45, 115.73, 116.36, 127.44, 130.26, 130.59, 133.05, 133.86, 135.63, 151.98, 152.23, 153.45, 153.45, 160.13, 165.79 (acetamide C=O), 168.26 (C=O). For C₂₅H₂₂FN₅O₃S calculated: 61.09 % C, 4.51 % H, 14.25 % N; found: 61.11 % C, 4.52 % H, 14.21 % N. HRMS [M-H]⁻: calculated: 490.1355, experimental: 490.1401.

Activity Studies

Matrix Metalloproteinase (MMP) Inhibition Assays

MMP-9, colorimetric kits were purchased from Enzo Life Sciences Inc. (Farmingdale, New York, NY, USA). The MMP Colorimetric Drug Discovery Kits are a complete assay system designed to screen MMP inhibitors using a thiopeptide as a chromogenic substrate (Ac-PLG-[2-mercapto-4-methyl-pentanoyl]-LG-OC₂H₅). The MMP cleavage site peptide bond is replaced by a thioester bond in the thiopeptide. Hydrolysis of this bond by an MMP produces a sulfhydryl group, which reacts with DTNB [5,5'-dithiobis(2-nitrobenzoic acid), Ellman’s reagent] to form 2-nitro-5-thiobenzoic acid, which can be detected by its absorbance at 412 nm. The assays were conducted in triplicate. The UV absorbance was read at 412 nm using a microplate reader (BioTek, PowerWave, Gen5 software, Winooski, VT, USA) at room temperature. NNGH was used as a control inhibitor.¹⁶ Data were expressed as Mean±SD.

The inhibitor% remaining activity of MMPs was calculated using the following equation:

Inhibitor% activity remaining = (V inhibitor/V control) × 100.

The inhibition (percent) of MMPs was calculated using the following equation:

I (%) = 100 – Inhibitor% activity remaining

Docking Studies

The crystal structure of MMP-9 was retrieved from the Protein Data Bank server (PDB code: 5I12). The protein preparation process, ligand preparation process, grid generation, docking and visualization studies were performed on Schrodinger’s Maestro molecular modeling package.¹⁷
The water molecules were removed from the crystal structure. Ligands were set to the physiological pH (pH = 7.4) at the protonation step. In molecular docking simulations: Glide/XP docking protocols were applied for the prediction of topologies of 5f, 5g and 5h at the active site of target structure, then they were docked to the active site of 5I12.

RESULTS and DISCUSSION

Chemistry

In this study, we aimed to synthesize novel 3,4,5-trisubstituted triazole derivatives. In the first of five steps, N-(4-hydroxyphenyl)acetamide and ethyl chloroacetate were refluxed in acetone.

After the reaction was completed, the acetone was evaporated, and the material was washed with water to acquire the intermediate. The obtained product, ethyl 2-(4-acetamidophenoxy)acetate (1) was reacted with hydrazine monohydrate in ethanol. The end of the reaction was checked by TLC, hydrazide compound was gained by filtration. N-(4-(2-oxo-2-(2-(phenylcarbamothioyl)hydrazineyl)ethoxy)phenyl)acetamide (2) was dissolved in ethanol and boiled with phenylisothiocyanate to synthesize N-phenyl-2-(2-(quinolin-8-yloxy)acetyl)hydrazine-1-carbothioamide (3) and then in next step this compound was refluxed with 2N potassium hydroxide prepared in ethanol. The reaction was terminated by controlling the TLC. The pH was set to 7 to allow the material to settle in a cold environment. At last, the resulting triazole molecules (4) were acquired with the reaction of appropriate 2-chloro-N-substituted acetamide derivatives to get the final nine molecules (5a-i) (Scheme 1). The target compounds (5a-i) were obtained purely and the structures of the compounds were elucidated with spectroscopic methods.
Scheme 1. Synthesis of the target compounds. Reaction conditions. a) Acetone, K$_2$CO$_3$, reflux, 6 h; b) EtOH, H$_2$NNH$_2$, H$_2$O, rt, 12 h; c) EtOH, reflux, 3 h; d) 2N KOH (ethanolic), reflux, 2 h; e) Acetone, K$_2$CO$_3$, rt, 4-5 h.

The structures of the synthesized materials were elucidated by $^1$H-NMR, $^{13}$C-NMR, HRMS and elemental analysis. The ppm values of the peaks of the -NH proton were detected at 9.78-12.80 ppm. The values of the proton peaks of O-CH$_2$ were seen at 5.03 ppm exactly and values of S-CH$_2$ protons were assigned at 4.17-4.33 ppm. The peak value of the CH$_3$ for all molecules was singlet at 1.97-2.00 ppm. For compound 5e, the substituent CH$_3$ peaks were observed at 1.98 and 2.40 ppm.

The O-CH$_3$ peak at 3.80 ppm in compound 5f was an expected value due to the substituent. The CH$_4$, O-CH$_2$-CH$_3$ and O-CH$_2$-CH$_3$ peaks attributed to the substituents in the compound 5g were observed at 1.97 ppm, 4.05 ppm, and 1.34 ppm.
ppm, respectively. Also, multiple peaks were observed in the aromatic region due to the phenyl ring in the substituent. In $^{13}$C-NMR spectra of the compounds, signals belong to C-CH$_3$ S-CH$_2$ and O-CH$_2$ were detected at 24.24-24.28 ppm, 36.00-36.54 ppm, and 60.51-60.56 ppm, respectively. The signal of carbonyl carbon was detected at 165.79-168.85 ppm in common with all final compounds. HRMS results of the compounds were processed in negative ion mode and peaks were detected following the molecular weights of the compounds.

**ADME Parameters**

Some properties of the synthesized compounds were displayed in Table 1, which were calculated by SwissADME.$^{18-20}$ The number of HBA was calculated minimum 6 and maximum 8, and also number of HBD was 2. Log P values were estimated between 2.23 and 3.65. All compounds have violations of RoF equal to or less than 2. Therefore, the final compounds may be applied for oral use according to forecast.$^{21}$ These findings were also in harmony with the activity results. Especially, compound 5e stand out via TPSA (161.10 Å$^2$) and Log P (3.36). These differences between 5e and other derivatives may be the reasons which explain the incoherent in the activity tests due the solubility.

**Table 1.** Some properties of synthesized compounds

<table>
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<th>HBA</th>
<th>HBD</th>
<th>TPSA</th>
<th>Log P$_{o/w}$</th>
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<td>2.92</td>
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<td>-3.76</td>
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</table>

**HBA:** H-bond acceptor, **HBD:** H-bond donor, **TPSA:** Topologic polar surface area (Å$^2$) **Log P$_{o/w}$:** Consensus Log P$_{o/w}$ (Average of all five predictions), **Log S:** Water Solubility, **RoF (V):** Rule of Five (violation number), **SD-1:** Standard Drug (N-Isobutyl-N-(4-methoxyphenylsulfonyl)glycyl hydroxamic acid).
Activity Results

All compounds (5a-5i) and the standard drug were investigated for the MMP-9 inhibition activity were shown in Table 2.

Table 2. Inhibition % on MMP-9

<table>
<thead>
<tr>
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<th>Inhibition %</th>
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<tr>
<td>5a</td>
<td>40.86 ± 1.62</td>
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<tr>
<td>5b</td>
<td>48.21 ± 1.27</td>
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<tr>
<td>5c</td>
<td>23.22 ± 3.57</td>
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<tr>
<td>5d</td>
<td>9.52 ± 2.06</td>
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<tr>
<td>5e</td>
<td>62.5 ± 2.53</td>
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<tr>
<td>5f</td>
<td>27.50 ± 1.52</td>
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<tr>
<td>5g</td>
<td>51.19 ± 2.06</td>
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<tr>
<td>5h</td>
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<tr>
<td>5i</td>
<td>41.07 ± 2.52</td>
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<tr>
<td>NNGH</td>
<td>91.10 ± 2.50</td>
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</table>

100 μg/mL of each synthesized compounds was used to determine the inhibition % on MMP-9. NNGH: N-Isobutyl-N-(4-methoxyphenylsulfonyl)glycyl hydroxamic acid, and 1.3 μM/mL of it was used to determine the inhibition% on MMP-9.

Enzyme Studies

According to the MMP-9 inhibition results, the most active compound was compound 5e with methyl substitution on benzothiazole ring. Also, compound 5g (6-ethoxybenzothiazole) showed moderate inhibition activity. These results were worthy of note that the development of new treatments against cancers like lung carcinoma. To better understand the structure-activity relationship, in our investigation we proceeded to the next step: molecular docking study.

Molecular Docking and Structure-Activity Relationship (SAR) Studies

For comparing the active and non-active/low effective structures, 5d, 5e, and 5g were docked to MMP-9 enzyme (PDB ID: 5I12). The docking poses collected as shown in Figure 1-6.
Figure 1. 3D best docking poses of compounds. [5d (green carbons), 5e (maroon carbons) and 5g (orange carbons) with Zn\(^{2+}\) (violet ball) in the active site of 5I12. The binding site amino acids were colored with turquoise blue carbons.]

Figure 2. 3D best docking pose of compounds 5d in active site of 5I12
**Figure 3.** 3D best docking pose of compound 5e in active site of 5I12

**Figure 4.** 2D interaction diagram of compound 5e in active site of 5I12
According to docking results, compound 5e interacted with His226, His236, Tyr245, Pro246, Tyr248, and Arg249 active site amino acids and Zn301 metal ion. Between the phenyl rings and the histamine amino acids occurred π-π stacking, and four H-bonds were viewed with the benzothiazole nitrogen, acetamide nitrogen, acetanilide nitrogen and oxygen. These bonds interacted with hydrophobic (Tyr and Pro) and positively charged amino acid (Arg). There was a π-cation interaction between triazole’s phenyl and Zn301.
Compound 5g interacted with Gly186, Leu188, His226, His230 amino acids, and Zn301 metal ion. Between phenyl and His226, and benzothiazole and His230 occurred π-π stacking. There was two H-bond displayed between the triazole nitrogen (3rd) and leucine, and acetonilide oxygen and glycine. Also, there was observed the salt bridge between Zn301 and acetamide nitrogen. Also, the π-cation interaction between the metal ion and triazole phenyl was observed.

Compound 5d interacted with Ala191, His226, His236, and Try248 amino acids and Zn301 metal ion. Between the phenyl rings and histamine amino acids (226, 236) occurred π-π stacking. There were two H-bonds which were between acetonilide nitrogen and Ala191, and acetamide nitrogen and Tyr248. There was observed π-cation (with phenyl of triazole) and metal coordination interactions.

Previous studies mentioned that zinc ion at the active site of the catalytic domain is closely related to the hydrolytic activity of MMP families.22,23 Thus, the effectiveness depends not only on the number of hydrogen bond but also on the bonds that the ligand forms with histamine and zinc. Our docking results point out that zinc ion, and its bonding amino acids (His226, His230, and His336) are the important residues as much as H-bonds in MMP-9 activity cavity. However, the relationship between the differences in inhibition% was found related to hydrogen bonds. The results of the docking study and in vitro enzyme study were in good harmony.

For less or non-active compounds, the electron-withdrawing groups at the benzothiazole ring decreased the inhibition activity. It may be caused by bulky atoms and/or non-hydrophobic affinity. Additionally, although there was not enough data to compare the electron-withdrawing or donating groups at phenyl, the enzyme study proved that the activities of the electron-withdrawing groups (-NO₂ and -F) showed that less than 50% inhibition. Therefore, for future studies, the electron-donating group and/or hydrophilic moiety may be thought. Also, as a new perspective, N substitution on the triazole ring may have a short carbon chain with composing π interactions for hydrophobicity and easy placement. The importance of π–π stacking between the ligand and the imidazole ring of histamine was underlined.22 On the other hand, the docking results showed that the substitutions at benzothiazole should comprise of carbon chain without heteroatoms. Although the ethoxy derivative (5g) showed the inhibition activity, it was seen that the oxygen atom didn’t show H-bond interaction. The methoxy (5f) derivative also didn’t display the inhibition activity, hence the group at benzothiazole may be constituted of the carbon chain without heteroatoms.
Briefly, we concluded the results that although final compounds have good pharmacokinetic profiles, only two compounds (5e and 5g) showed an attractive activity. The distinctness between the active compounds and others were related to the hydrophobic properties. Also, the interactions between the ligand and enzyme were discussed for future studies. Hopefully, these modifications especially will be increased the anticancer activity through MMP-9 inhibition or at least give an option. In the next studies, these are the main data in the designing strategy.

CONFLICT OF INTEREST

The author confirms that this article content has no conflict of interest.
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14. Kaur, R., Ranjan Dwivedi, A., Kumar, B., & Kumar, V. (2016). Recent developments on 1, 2, 4-triazole nucleus in anticancer compounds: a review. *Anti-Cancer Agents in Medicinal Chemistry (Formerly Current Medicinal Chemistry-Anti-Cancer Agents)*, 16(4), 465-489.


