# Cytotoxic activity of *Achillea arabica* Kotschy against renal cancer cell lines

Güneş Güniz GÜNGÖR<sup>1</sup>, Fatma TOSUN<sup>2\*</sup>

1 Istanbul Medipol University, Graduate School of Health Sciences, Department of Pharmacognosy, Istanbul, Türkiye 2 Istanbul Medipol University, School of Pharmacy, Department of Pharmacognosy, Istanbul, Türkiye

#### ABSTRACT

Achillea arabica Kotschy, belonging to the Asteraceae family, is a plant whose biological activities, such as wound healing, antimicrobial, anticancer, and antioxidant properties, have been evidenced by studies. It has been investigated for its cytotoxic activity in various cell lines, including AGS, MCF7, SW742, SKLC6, A375, PLC/PRF/5, HT29, and HepG2. However, no study has been conducted on any cell line related to kidney cancer. In this study, the cytotoxic activities of the extracts prepared from the root and aerial parts of the plant with dichloromethane, ethyl acetate, and methanol were investigated against A498 and UO31 kidney cancer cell lines. The highest activity was observed in the dichloromethane extract of the aerial parts of the plant on both cancer cell lines. The dichloromethane extract of the aerial parts showed 63% inhibition on the A498 cell line and 56% inhibition on the UO31 cell line at a concentration of 25  $\mu$ g/mL. The results we have obtained have been of a quality that will lead to new research in this context.

Keywords: cytotoxic activity, Achillea arabica, renal cancer

#### INTRODUCTION

Cancer is one of the leading causes of death worldwide<sup>1</sup>. It is also a disease with typical properties such as abnormal cell growth, invasion, metastasis and mu-

<sup>\*</sup> Corresponding author: Fatma TOSUN E-mail: ftosun@medipol.edu.tr ORCIDs: Güneş Güniz GÜNGÖR: 0009-0002-0911-0946 Fatma TOSUN: 0000-0003-2533-5141 (Received 16 Jul 2024, Accepted 5 Sept 2024)

<sup>©</sup> Medipol University Press / ISSN: 2636-8552

tations<sup>2</sup>. According to the studies, 2,001.140 new cancer cases and 611,720 cancer deaths are expected to occur in 2024. The fact that most of the drugs in cancer treatment often have side effects such as ischemic heart disease, vomiting, bone marrow suppression, myelosuppression, liver dysfunction, fatigue and hypertension, which further hinder the treatment process, has led researchers to search for different treatments<sup>3</sup>. Therefore, the discovery of new treatment strategies or chemotherapeutics with minimal or no side effects that are easily accessible, cost-effective, and more effective for treating this deadly disease has become one of the major goals in cancer therapy<sup>4</sup>. In addition, chemotherapy treatment significantly reduces the patient's quality of life as it damages normal cells. Therefore, it is very important to search for new anti-tumor agents that are specific to tumor cells<sup>5</sup>.

Throughout the ages, it is known that people have utilized nature to meet their basic needs. This has also been the case for the use of natural products as medicines in the treatment of various diseases<sup>6</sup>. Plant extracts and plantderived natural compounds such as glycosides, alkaloids, tannins, terpenes, coumarins and flavonoids are known to inhibit the growth of cancer cells such as kidney, lung, breast and colorectal cancer cells<sup>7,8</sup>. Kidney cancer (Renal Cell Carcinoma-RCC) is the second most common type of cancer of the urinary system9. Currently, RCC constitutes approximately 2-3% of all adult malignancies worldwide and ranks as the 12th most common malignancy and the third most common urogenital cancer<sup>10</sup>. RCC has a high metastatic potential to the lung, liver, bone, head and neck11. Approximately 30% of RCC cases are diagnosed at an advanced or metastatic stage, and according to the International Metastatic Renal Cell Carcinoma Database Consortium (IMDC) criteria, nearly 80% of these patients belong to the intermediate or low risk group<sup>12</sup>. Due to limited treatment options, less than 40% of patients survive for  $\geq 5$  years after diagnosis. Additionally, the lack of routine adjuvant therapy in the clinic is a key reason for the recurrence of kidney cancer, as kidney cancer is resistant to both chemotherapy and radiotherapy. Therefore, there is a need to search for new compounds and develop targeted therapies for kidney cancer<sup>13</sup>.

The *Achillea* genus includes 138 accepted species distributed in Europe, Asia, North Africa, North and Central America, Alaska, and Greenland<sup>14</sup>. The genus *Achillea* is represented by 50 species in Turkey<sup>15</sup>. *Achillea* species were reported to contain polyphenols, flavonoids, phenolic and quinic acid derivatives, sesquiterpenes lactones and essential oils; and to have anticancer, antioxidant, antimicrobial, anti-inflammatory, analgesic, antipyretic, antidiabetic, antihelmintic and antihypertensive activities<sup>16-24</sup>. Previous studies have shown that the extracts of *Achillea arabica* have anticancer, antimicrobial, antioxidant, anti-inflammatory, anticholinesterase, analgesic, anxiolytic and wound-healing effects<sup>17,23-30</sup> and contain phenolic compounds, flavonoids, sesquiterpenes lactones and essential oils<sup>17,29-38</sup>.

When cytotoxic activity studies conducted on different cell lines were compiled to determine the anticancer efficacy of *A. arabica*, no study was found to be conducted on the A498 and UO31 kidney cancer cell lines.

### METHODOLOGY

### Plant material

*Achillea arabica* Kotschy used in this study was collected from Adana-Saimbeyli region on June 12, 2016. Herbarium specimens of the plant identified by Prof. Mecit Vural are registered in Istanbul University Faculty of Pharmacy Herbarium (ISTE 115056).

# Preparation of extracts

The extraction process was conducted using solvents of increasing polarity, as they selectively consume compounds with different chemical structures based on their polarities. Additionally, considering the possibility of degradation with heat, the extraction process was carried out at room temperature. The pulverized plant material was successively macerated with dichloromethane (DCM), ethyl acetate (EtOAc), and methanol (MeOH) at room temperature. After each extraction with a solvent, the plant material was dried to remove the solvent completely and then subjected to extraction with the next solvent. After consumption, the extracts filtered through filter paper were concentrated using a rotary evaporator at 40°C.

# Cytotoxicity assay on cancer cells

*In-vitro* 2Day XTT cytotoxic activity test<sup>39</sup> was applied to extracts of the roots and aerial parts. The 2Day XTT bioactivity assay is an *in-vitro* colorimetric cytotoxic activity test. XTT bioactivity assays were performed at the NCI MTP Experiment Development and Imaging Department. Kidney cancer cell lines (A498, UO31) were used in XTT cytotoxic activity tests. Sanguinarine chloride hydrate was used as a control in the experiment. The assay was performed as described previously<sup>8</sup>.

# **RESULTS and DISCUSSION**

In this study, the *Achillea arabica* plant was investigated for the first time in terms of cytotoxic activity on A498 and UO31 cell lines in kidney cancer. The

extract yields obtained by DCM, EtOAc and MeOH extraction of the aerial parts and roots of the plant are given in Table 1. The yield of the extract is also important for the calculation of the plant material to be used for the subsequent isolation of the active compounds.

As a result of the cytotoxic activity studies of the extracts obtained from the aerial parts and roots of the plant, the % inhibition values they showed on A498 and UO31 kidney cancer cell lines are given in Table 2.

	yield (a/a) %				
Extracts	Aerial parts	Roots			
1	1.32%	1.18%			
2	0.69%	0.68%			
3	3.64%	3.36%			

Table 1. % yield obtained by extraction of the aerial parts and roots of A. arabica

1: DCM extract; 2: EtOAc extract; 3: MeOH extract

	inhibition %						
	A498			U031			
Plant parts	1	2	3	1	2	3	
Roots	40	45	54	43	53	53	
Aerial parts	63	41	46	56	43	54	

1: DCM extract; 2: EtOAc extract; 3: MeOH extract

Cytotoxic activity was determined on the kidney cancer cell lines (A498 and UO31) by *in-vitro* 2Day XTT cytotoxic activity assay. On the A498 cancer cell line, the root MeOH extract, and the aerial parts DCM extract exhibited over 50% inhibition at a concentration of 25  $\mu$ g/mL, while on the UO31 cell line, the root EtOAc and MeOH extracts, as well as the aerial parts DCM and MeOH extracts, showed over 50% inhibition at a concentration of 25  $\mu$ g/mL. The highest activity on both cancer cell lines was observed with the DCM extract of the aerial parts. The DCM extract of the aerial parts exhibited 63% inhibition on the A498 cell line and 56% inhibition on the UO31 cell line at a concentration of 25  $\mu$ g/mL.

In recent years, the therapeutic potential of medicinal plant extracts in preventing and treating cancer has attracted scientists' attention. Approximately 75% of approved cancer drugs have been developed based on agents of natural source. Plant-derived active compounds support treatment at all stages of cancer and are multi-targeted and non-toxic<sup>40</sup>.

Previously, *A. arabica* has been tested against HeLa (cervical cancer), AGS (gastric adenocarcinoma), MCF7 (breast cancer), SW742 (colorectal adenocarcinoma) SKLC6 (lung cancer), A375 (skin cancer), PLC/PRF/5 (liver cancer)<sup>24,30</sup>. In another study, *A. arabica* (*A. biebersteinii*) extracts prepared with hexane, chloroform, and methanol were found to be effective in HT-29 (colorectal carcinoma) cell line, increased the activity of the 5-FU compound used in treatment, induced apoptosis by regulating PTEN/Akt/mTOR signaling pathway, and inhibited angiogenesis<sup>41</sup>. Ag-NPs were synthesized using *A. arabica* (*A. biebersteinii*) flower extract has been reported to induce apoptosis on MCF-7 cell line and can be considered a potential chemotherapeutic agent in treating breast cancer<sup>42</sup>. This study represents the first investigation on the cytotoxic activity of *A. arabica* on A498 and UO31 cell lines.

Previous studies reported that the extracts of the *Achillea* species exhibited cytotoxic activities against different cancer cell lines<sup>17-22,43,44</sup>, but there are no reports in the literature dealing with the cytotoxic activities of the *Achillea* species on the renal cancer cell lines. Therefore, our study is important in terms of being the first cytotoxic activity study on the kidney cancer cell lines not only on the *A. arabica* species but also on the *Achillea* genus and contributed to the literature on this regard.

In conclusion, bioactivity-guided fractionation of the DCM extract of the aerial parts, exhibited the highest activity on both cancer cell lines, is planned to isolate and identify the compounds responsible for the activity.

#### STATEMENT OF ETHICS

There is no ethical statement provided.

#### CONFLICT OF INTEREST STATEMENT

The authors state that they have no conflicts of interest.

#### AUTHOR CONTRIBUTIONS

These authors contributed to the work equally.

#### FUNDING SOURCES

There are no sources of funding indicated.

#### ACKNOWLEDGMENTS

We thank Dr. John A. Beutler, Molecular Targets Laboratory, CCR, NCI, Frederick, MD, U.S.A., for the cytotoxic activity testing.

We thank Prof. M. Vural for identifying of plant material.

#### REFERENCES

1. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. CA Cancer J Clin, 2022;72(1):7-33. Doi: 10.3322/caac.21708

2. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin, 2018;68(6):394-424. Doi: 10.3322/caac.21492

3. Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. CA Cancer J Clin, 2024;74(1):12-49. Doi: 10.3322/caac.21820

4. Banik K, Khatoon E, Harsha C, Rana V, Parama D, Thakur KK, et al. Wogonin and its analogs for the prevention and treatment of cancer: a systematic review. Phytother Res, 2022;36(5):1854-1883. Doi: 10.1002/ptr.7386

5. Deb DD, Parimala G, Devi SS, Chakraborty T. Effect of thymol on peripheral blood mononuclear cell PBMC and acute promyelotic cancer cell line HL-60. Chem Biol Interact, 2011;193(1):97-106. Doi: 10.1016/j.cbi.2011.05.009

6. Majolo F, Delwing LKDOB, Marmitt DJ, Bustamante-Filho IC, Goettert MI. Medicinal plants and bioactive natural compounds for cancer treatment: important advances for drug discovery. Phytochem Lett, 2019;31:196-207. Doi: 10.1016/j.phytol.2019.04.003

7. Isyaka SM, Mas-Claret E, Langat MK, Hodges T, Selway B, Mbala BM. Cytotoxic diterpenoids from the leaves and stem bark of *Croton haumanianus* (Euphorbiaceae). Phytochem, 2020;178:112455. Doi: 10.1016/j.phytochem.2020.112455

8. Tosun F, Aytar EC, Beutler JA, Wilson JA, Miski M. Cytotoxic sesquiterpene coumarins from the roots of *Heptaptera cilicica*. Rec Nat Prod, 2021;15(6):529-536. Doi: 10.25135/rnp.242.21.02.1990

9. Mihoğlugil F, Akalgan D, Tosun F. Cytotoxicity screening of some Turkish plants against renal cancer cells. J Res Pharm, 2023;27(2):636-641. Doi: 10.29228/jrp.346

10. Yu X, Du Z, Zhu P, Liao B. Diagnostic, prognostic, and therapeutic potential of exosomal microRNAs in renal cancer. Pharmaco Rep, 2024;76:273-286. Doi: 10.1007/s43440-024-00568-7

11. Kweon HT, Yoo JS, Hong, YT. Tongue metastasis from renal cell carcinoma: a rare case presentation. Ear Nose Throat J, 2024;1-3. Doi: 10.1177/01455613231226038

12. Lalani AKA, Heng D, Basappa NS, Wood L, Iqbal N, McLeod D. Evolving landscape of firstline combination therapy in advanced renal cancer: a systematic review. Ther Adv Med Oncol, 2022;14:1-17. Doi: 10.1177/17588359221108685

13. Zhu C, Na N, Sheng H, Feng B, Wang H, Zhu P. Ginkgolic acid inhibits the growth of renal cell carcinoma cells via inactivation of the EGFR signaling pathway. Exp Ther Med, 2020;19(4):2949-2956. Doi: 10.3892/etm.2020.8570

14. Plants of the World Online. *Achillea arabica* Kotschy [Internet]. Kew Royal Botanic Gardens [Jul 16, 2024]. Available from: https://powo.science.kew.org/taxon/urn:lsid:ipni. org:names:173817-1/

15. Güner A, Aslan S, Ekim T, Vural M, Babaç MT, editors. Türkiye bitkileri listesi (damarlı bitkiler). İstanbul: Nezahat Gökyiğit Botanik Bahçesi ve Flora Araştırmaları Derneği Yayını; 2012.

16. Saeidnia S, Gohari AR, Mokhber-Dezfuli N, Kiuchi FA. Review on phytochemistry and medicinal properties of the genus *Achillea*. DARU: J Pharm Sci, 2011;19(3):173-186. 17. Barda C, Grafakou ME, Tomou EM, Skaltsa H. Phytochemistry and evidence-based traditional uses of the genus *Achillea* L.: an update (2011–2021). Sci Pharm, 2021;89(4):50-86. Doi: 10.3390/scipharm89040050

18. Awad BM, Habib ES, Ibrahim AK, Wanas AS, Radwan MM, Helal MA, et al. Cytotoxic activity evaluation and molecular docking study of phenolic derivatives from *Achillea fragrantissima* (Forssk.) growing in Egypt. Med Chem Res, 2017;26:2065-2073. Doi: 10.1007/s00044-017-1918-6

19. Tsiftsoglou OS, Krigas N, Gounaris C, Papitsa C, Nanouli M, Vartholomatos E, et al. Isolation of secondary metabolites from *Achillea grandifolia* Friv. (Asteraceae) and main compounds' effects on a glioblastoma cellular model. Pharmaceutics, 2023;15(5):1383-1397. Doi: 10.3390/ pharmaceutics15051383

20. Mohammed HA, Abd-Elraouf M, Sulaiman GM, Almahmoud SA, Hamada FA, Khan RA, et al. Variability in the volatile constituents and biological activities of *Achillea millefolium* L. essential oils obtained from different plant parts and by different solvents. Arab J Chem, 2023;16(9):1-15. Doi: 10.1016/j.arabjc.2023.105103

21. Taşkın D, Doğan M, Ermanoğlu M, Ermanoglu M, Arabaci T. Achillea goniocephala extract loaded into nanochitosan: in vitro cytotoxic and antioxidant activity. Clin Exp Health Sci, 2021;11(4):659-666. Doi: 10.33808/clinexphealthsci.972180

22. Eruygur N, Ataş M, Tekin M, Cevik, O. Evaluation of in vitro antioxidant, antimicrobial and cytotoxic activities of crude ethanol extract and fractions of Achillea sintenisii Hub. Mor. Clin Exp Health Sci, 2023; 13(3):517-524. Doi: 10.33808/clinexphealthsci.1058614

23. Raudone L, Radušiene J, Seyis F, Yayla F, Vilkickyte G, Marksa M, et al. Distribution of phenolic compounds and antioxidant activity in plant parts and populations of seven underutilized wild *Achillea* species. Plants, 2022;11(3):447-466. Doi: 10.3390/plants11030447

24. Yilmaz MA, Ertaş A, Yener İ, Türkmenoğlu FP, Ölmez ÖT, Öztürk M, et al. Chemical fingerprints and bioactivities of 12 Anatolian Achillea species by LC-MS/MS with chemometric approach: novel phytonutrients, natural food preservatives and chlorogenic acid sources. Turk J Bot, 2022;46(5):473-489. Doi: 10.55730/1300-008X.2723

25. Toplan GG, Taşkın T, İşcan G, Göger F, Kürkçüoğlu M, Civaş A, et al. Comparative studies on essential oil and phenolic content with in vitro antioxidant, anticholinesterase, antimicrobial activities of *Achillea biebersteinii* Afan. and *A. millefolium* subsp. *millefolium* Afan. L. growing in Eastern Turkey. Molecules, 2022;27(6):1956. Doi: 10.3390/molecules27061956

26. Bashi DS, Fazly Bazzaz BS, Sahebkar A, Karimkhani MM, Ahmadi A. Investigation of optimal extraction, antioxidant, and antimicrobial activities of *Achillea biebersteinii* and *A. wilhelmsii*. Pharm Biol, 2012;50(9):1168-1176. Doi: 10.3109/13880209.2012.662235

27. Akkol EK, Koca U, Pesin I, Yilmazer D. Evaluation of the wound healing potential of *Achillea biebersteinii* Afan. (Asteraceae) by *in vivo* excision and incision models. eCAM, 2011;1-7. Doi: 10.1093/ecam/nep039

28. Varasteh-Kojourian M, Abrishamchi P, Matin MM, Asili J, Ejtehadi H, Khosravitabar F. Antioxidant, cytotoxic and DNA protective properties of *Achillea eriophora* DC. and *Achillea biebersteinii* Afan. extracts: a comparative study. Avicenna J Phytomed, 2017;7(2):157-168.

29. Bariş Ö, Güllüce M, Şahin F, Özer H, Kiliç H, Özkan H, et al. Biological activities of the essential oil and methanol extract of Achillea biebersteinii Afan. (Asteraceae). Turk J Biol, 2006;30(2):65-73. Doi: 10.5897/JMPR10.560

30. Ghavami G, Sardari S, Shokrgozar MA. Anticancerous potentials of *Achillea* species against selected cell lines. J Med Plants Res, 2010;4(22):2411-2417. Doi: 10.5897/JMPR10.560

31. Al-Shuneigat JM, Al-Sarayreh SA, Al-Qudah MA, Al-Saraireh YM. Antibacterial and antibiofilm activity of essential oil of *Achillea biebersteinii* and its mode of action. J Pharm Pharmacogn Res, 2020;8(2):155-166.

32. Cirak C, Radusiene J, Raudone L, Vilkickyte G, Seyis F, Marksa M, Yayla F. Phenolic compounds and antioxidant activity of *Achillea arabica* populations. S Afr J Bot, 2022;147:425-433. Doi: 10.1016/j.sajb.2022.02.006

33. Oskay E, Yesilada A. Four flavonoids and three other constituents from *Achillea biebersteinii*. J Nat Prod, 1984;47(4):742-742. Doi: 10.1021/np50034a041

34. Abd-Alla H, Shalaby N, Hamed M, El-Rigal NS, Al-Ghamdi S, Bouajila J. Phytochemical composition, protective and therapeutic effect on gastric ulcer and a-amylase inhibitory activity of *Achillea biebersteinii* Afan. Arch Pharm Res, 2015;39:10-20. Doi: 10.1007/S12272-014-0544-9

35. Şabanoğlu S, Gökbulut A, Altun ML. Characterization of phenolic compounds, total phenolic content and antioxidant activity of three *Achillea* species. J Res Pharm, 2019;23(3):567-576. Doi: 10.12991/jrp.2019.164

36. Gawel-Beben K, Strzepek-Gomółka M, Czop M, Sakipova Z, Głowniak K, Kukula-Koch W. *Achillea millefolium* L. and *Achillea biebersteinii* Afan. hydroglycolic extracts-bioactive ingredients for cosmetic use. Molecules, 2020;24:3368. Doi: 10.3390/molecules25153368

37. Chialva F, Monguzzi F, Manitto P, Akgül, A. Essential oil constituents of *Achillea biebersteinii* Afan. J Essent Oil Res, 1993;5(1):87-88. Doi: 10.1080/10412905.1993.9698176

38. Tosun F, Kürkçüoğlu M. The essential oils of two Achillea L. species from Turkey. Acta Pharm Sci, 2018;56(2):59-66. Doi: 10.23893/1307-2080.aps.05611

39. Devkota KP, Covell D, Ransom T, McMahon JB, Beutler JA. Growth inhibition of human colon carcinoma cells by sesquiterpenoids and tetralones of *Zygogynum calothyrsum*. J Nat Prod, 2013;76:710-714. Doi: 10.1021/np400042q

40. Wani ZA, Guru SK, Rao AS, Sharma S, Mahajan G, Behl A. A novel quinazolinone chalcone derivative induces mitochondrial dependent apoptosis and inhibits PI3K/Akt/mTOR signaling pathway in human colon cancer HCT-116 cells. Food Chem Toxiol, 2016;87:1-11. Doi: 10.1016/j. fct.2015.11.016

41. Erdoğan MK, Ağca CA, Aşkın H. *Achillea biebersteinii* extracts suppress angiogenesis and enhance sensitivity to 5-fluorouracil of human colon cancer cells via the PTEN/AKT/mTOR pathway *in vitro*. Asian Pac J Trop Biomed, 2020;10(11):505-515. Doi: 10.4103/2221-1691.294091

42. Baharara J, Namvar F, Ramezani T, Mousavi M, Mohamad R. Silver nanoparticles biosynthesized using *Achillea biebersteinii* flower extract: apoptosis induction in MCF-7 cells via caspase activation and regulation of Bax and Bcl-2 gene expression. Molecules, 2015;20(2):2693-2706. Doi: 10.3390/molecules20022693

43. Boutennoun H, Boussouf L, Rawashdeh A, Al-Qaoud K, Abdelhafez S, Kebieche M, et al. *In vitro* cytotoxic and antioxidant activities of phenolic components of Algerian *Achillea odorata* leaves. Arab J Chem, 2017;10(3):403-409. Doi: 10.1016/j.arabjc.2014.05.013

44. Csupor-Loffler B, Hajdu Z, Zupko I, Rethy B, Falkay G, Forgo P, et al. Antiproliferative effect of flavonoids and sesquiterpenoids from *Achillea millefolium* on cultured human tumour cell lines. Phytother Res, 2009;23(5):672-676. Doi: 10.1002/ptr.2697