HPLC Assay for Aflatoxins in Dried Red Peppers and Feedstuffs in Turkey

Türkiye'de Kurutulmuş Kırmızı Biberlerde ve Yemlerde HPLC ile Aflatoksin Aranması

Gülden Z. Omurtag¹, Gülin Atak¹, Gülçin Keskin², Ömer Ersoy¹

- Marmara University, Faculty of Pharmacy, Department of Pharmaceutical Toxicology, 81010, Haydarpaşa-İstanbul, Turkey.
- ² ALKE, Department of Research and Development, Çınardere Mah. 3. Küme Sok. No. 18, Dolayoba-Pendik, İstanbul, Turkey.

Abstract

The purpose of this study was to investigate aflatoxins (AFs) B_1 , B_2 , G_1 and G_2 in contaminated dried red peppers and feedstuffs consumed in Turkey. The AFs were detected using high performance liquid chromatography (HPLC) and the results higher than 1 ppb were further confirmed by thin layer chromatography (TLC). In HPLC, the detection limit of B_1 and G_1 was 0.075 ng and for B_2 and G_2 it was 0.060 ng. In TLC, the detection limit of AFs was 1 ng. A total of 26 commercially available dried red pepper and 53 feedstuff specimens were analyzed. In Turkey for spices the allowed upper B_1 level is 5 ppb and for all foods total AF level is 10 ppb. In dried red peppers, the B_1 level higher than 5 ppb

was 42.3% and the total AF level greater than 10 ppb was 38.4%. In Turkey for mixed feedstuffs the allowed upper total AF level is 50 ppb and in 7.5% of the feedstuffs the total level was higher than 50 ppb.

Key Words: Aflatoxin; feedstuff; dried red pepper; HPLC.

Introduction

Aflatoxins (AFs) are highly toxic, carcinogenic compounds that are of major concern as a toxic contaminant in foods, feedstuffs, herbs and spices. It is believed that the sun-drying process of spices presents the greatest potential for AF contamination. The AFs are produced on agricultural commodities in the field under stress condition or when high moisture and warm temperature exist in storage (Holcomb et al., 1992). "Turkey-X disease" was the outbreak first in the UK and these compounds were discovered in the 1960s (Macdonald and Castle, 1996). AF B₁, B₂, G₁ and G₂ are acutely carcinogenic metabolites of the Aspergillus flavus and Aspergillus parasiticus (Scott, 1978). The International Agency for Research on Cancer has placed B₁ on their list of probable human carcinogen (WHO, 1987). Toxicologically, AF may be regarded as a quadruple threat: it can function as a potent toxin, a carcinogen, a teratogen, and a mutagen (Ciegler, 1975). Epidemiological studies have shown a correlation between AF exposure and primary hepatocellular carcinoma incidence in several Third World Countries and people in these regions suffer a high incidence of liver cancer. After metabolic activation of B₁, it is converted to the B₁ 8,9-epoxide, the ultimate carcinogen (Garner et al., 1993; Harrison et al., 1993). B₁ single dose (per os) is acutely toxic in all species studied, for example with an LD₅₀ ranging from 0.3 mg.kg⁻¹ for the duckling to 9.0 mg.kg⁻¹ for the mouse (Ciegler, 1975).

It was reported that in some samples of spices, especially chilli powder, over 20 ppb total AFs were found and more than 50% of the spice samples were contaminated at levels greater than 1 ppb (Garner *et al.*, 1993). In 20% of the 10 red pepper samples collected in Tokyo, the average B₁, B₂, G₁ amounts detected were 5.9, 0.2 and 0.2 ppb, respectively (Tabata *et al.*, 1993). In a study of 157 retail samples in the UK which included curry powders, pepper,

cayenne pepper, chilli, paprika, ginger, cinnamon and coriander it was found that nearly 95% of samples contained below 10 ppb total AFs and only nine samples had higher levels with a maximum concentration of 48 ppb in one chilli powder sample (Macdonald and Castle, 1996). About 8700 ppb AF was found in mouldy corn in the USA (Cavalheiro, 1983). The Moroccan poultry feeds were surveyed from 1989 to 1991 and the levels of AFB1 contamination ranged from 20 to 200 ppb except for four samples which contained higher levels of B1 as 2000-5625 ppb (Kichou and Walser, 1993). In Cyprus, AF contamination in peanut butter was found as 56.7% and the highest level of AFB1 was 700 ppb (Ioannou-Kakouri *et al.*, 1999). Various methods have been used for the determination of AFs based on HPLC (Dunne *et al.*, 1993; Garner *et al.*, 1993; Holcomb *et al.*, 1992; Howell and Taylor, 1981; Paulsch *et al.*, 1988), gas chromatography-mass spectrophotometry (GC/MS) (Holcomb *et al.*, 1992), TLC (Ehrlich and Lee, 1984; Majerus and Zakaria, 1992; Nesheim *et al.*, 1999), HPTLC (Ioannou-Kakouri *et al.*, 1999), enzyme-linked immunosorbant assay (ELISA) (Adachi *et al.*, 1991; Kichou and Walser, 1993), immunoaffinity (Trucksess *et al.*, 1990), mini column chromatography (Arim *et al.*, 1999).

In many countries the upper limits for the AF levels as ppb in foods is as; 20 ppb in the USA, 15 ppb in Canada, 10 ppb in Japan, France and UK, 5 ppb in Australia (Tabata *et al.*, 1993). In the USA, concentrations of up to 20 ppb are permitted in feedstuffs (Cavalheiro, 1983). Italy established maximum tolerated levels in spices as 20 ppb (FAO, 1997). The AF, T-2 toxin and fumonisin B₁, B₂, are all mycotoxins, yet in Turkey the guideline appear only for AF (Omurtag *et al.*, 1998; Omurtag, 2001; Omurtag and Yazıcıoğlu, 2000; Omurtag and Yazıcıoğlu, 2001). The upper limits are as 5 ppb AFB₁ in spices/all foods and 10 ppb total AF in all foods (T.C. Resmi Gazete, 1997). The upper limit in mixed feedstuffs is 50 ppb for total AF (T.C. Resmi Gazete, 1990). The AF problem in Turkey was encountered first in 1967 with hazelnut and later in 1972 and 1974 with pistachio and dried fig and in 1994 with red peppers (Çamlıbel, 1995). The purpose of this study was to determine by HPLC the extend of AF in dried red peppers and feedstuffs consumed in Turkey.

Materials and Methods

Materials: Total number of samples was 79 of which 26 were dried red peppers and 53 were feedstuffs. All samples were collected from various cities of Turkey. The collected dried red pepper samples were powdered. Each feedstuff sample was blended and finely ground using an Erweka mill (AR-400 TG2S-57222) with Retsch Test Sieve (Serial number 703429, mesh 20). Using this sieve mixture a 250 g subsample was obtained (stored – 20° C) and finally a 50 g was taken from it. The sample preparation, HPLC and TLC procedures were reported elsewhere (Omurtag *et al.*, 1998).

Chemicals: AF B_1 , B_2 , G_1 and G_2 (Sigma A-6636, 9887, 0138 and 0263 respectively) were used as standards. B_1 , B_2 , G_1 and G_2 were dissolved in benzene-acetonitrile (98:2 v/v) so as to contain 20 ng of AF.mL⁻¹ for HPLC and 10 μ g of AF.mL⁻¹ for TLC. The SPE (Solid-phase extraction) column was a silica cartridge (JT Baker, 7086-07) and other chemicals were of HPLC-grade Merck products. Bidistilled water was used.

Sample preparation: 50 g of sample was mixed in a Waring Blender with 200 mL of methanol-water (85:15, v/v) for 3 min. After filtration, 40 mL of 10% sodium chloride solution was added to 40 mL of filtrate and this solution was extracted with 2×25 mL of hexane. The hexane extracts were discarded and the defatted solution was extracted with 2×25 mL of chloroform and organic phase evaporated. The SPE column was conditioned with 3-mL hexane followed by 3-mL diethylether, and 3 mL methylene chloride. AFs were eluted with 6-mL chloroform-acetone (9:1 v/v). The eluates were collected into 10mL screw cap vials and evaporated to dryness under a stream of nitrogen.

Procedure for the HPLC: A volume of 0.2 mL of hexane was added to the residue, vortexed and 0.2 mL of 80 % trifluoroacetic acid (TFA) was added and revortexed. 2.3 mL of CH₃CN- 2.5% acetic acid (10:90, v/v) was added and mixed thoroughly; then the lower (aqueous) layer was filtered through a Millex HV filter (0:45 μ m). 25 μ L and/or 50 μ L of this sample was injected into the HPLC column. Samples were prepared and analysed in triplicate. The areas of the peaks were used for quantification.

Apparatus: The HPLC (Waters Corp., U.S.A.) was used with a Model 510 Pump and a M 420-AC Fluorescence Detector (Waters). Fluorescence was recorded at excitation (λ_{Ex}) and emission (λ_{Em}) wavelengths of 365 and 425 nm, respectively. The injector was a Rheodyne 7725 sample injector (100 μ L accessory). A reverse phase, μ Bondapak C₁₈ (10 μ m) (Waters), 3.9×300 mm I.D. analytical column was protected by a C₁₈ guard column. The data station was an Unicam 4880 Chromatography Data Handling System.

Operating conditions: The mobile phase was methanol-acetonitrile-5.0 % acetic acid (14:14:72, v/v/v); flow rate was 1.7 mL.min⁻¹; chart speed was 0.25 cm.min⁻¹ and column temperature was ambient. The mobile phase was filtered through a Millipore HV (0.45 μ m) membrane filter and sonicated. An AF (B₁+B₂+G₁+G₂) mixed standard (0.02 ng. μ L⁻¹) was derivatisated with TFA. An aliquot (25 and/or 50 μ L) injected to the HPLC column and linear regression lines for B₁, B₂, G₁ and G₂ were found to be y=2.6325x-33.3 (r=0.9984) y=3.3825x+21 (r=0.9998) y=3.0375x-24 (r=0.9984) and y=3.57x-18.7 (r=0.9998), respectively. The retention times of B₁, B₂, G₁ and G₂ were 5.65 min, 10.90 min, 4.31 min, and 7.76 min, respectively.

Results and Discussion

All the specimens were analyzed by HPLC and TLC was used to verify the positive results higher than 1 ppb. The B₁ detected in dried red peppers and feedstuffs were 65% and 17%, respectively and the complete list of the specimen results with the mean values of AFs are given in the Table 1. HPLC chromatograms of AFs standards and one dried red pepper sample is given in Figure 1. The highest level of B₁ in dried red peppers and feedstuffs were 56 ppb and 59.9 ppb, respectively. In dried red peppers the B₁ level higher than 5 ppb was 42.3% and the total AFs level higher than 10 ppb was 38.4% of the total specimens. In Turkey for mixed feedstuffs the allowed upper total AF level is 50 ppb and in 7.5% of the feedstuffs this value was exceeded. In 1997, 32 dried red pepper samples were collected in Turkey, the detected highest B₁ level was 109.7 ppb and about 50% of the samples B₁ level

was higher than 5 ppb (Omurtag *et al.*, 1998). Multi-mycotoxin residue analysis by TLC with fluorodensitometric procedures was made in totally 302 feeds and feedstuffs obtained from 25 feed factories in Turkey and incidences of AFB₁, G₁, G₂ were 11.2%, 1.98% and 0.3%, respectively (Özkazanç *et al.*, 1992).

In this study, the detection limits of HPLC were twice the signal-to-noise ratio, 0.075 ng for B_1 and G_1 and 0.06 ng for B_2 and G_2 . The detection limits of TLC were 1 ng for AFs. The HPLC recoveries of B_1 , B_2 , G_1 and G_2 from 4 ppb spiked AF-free dried red peppers and feedstuffs were 91±1.79%, 71±3.16%, 97±2.28%, 67±2.37% and 82±6.24%, 74±4.09%, 82±4.42% and 65±5.03%, respectively. The recovery results were expressed as the mean of five different manipulations including the extraction from the same AF-free specimen.

For protection of food supplies from mycotoxin contamination, special care must be given from the earliest point of production until consumption. As a consequence, the exposure to AFs will not only put some additional health risks to the consumers, but also arises to be an economic problem for the producers.

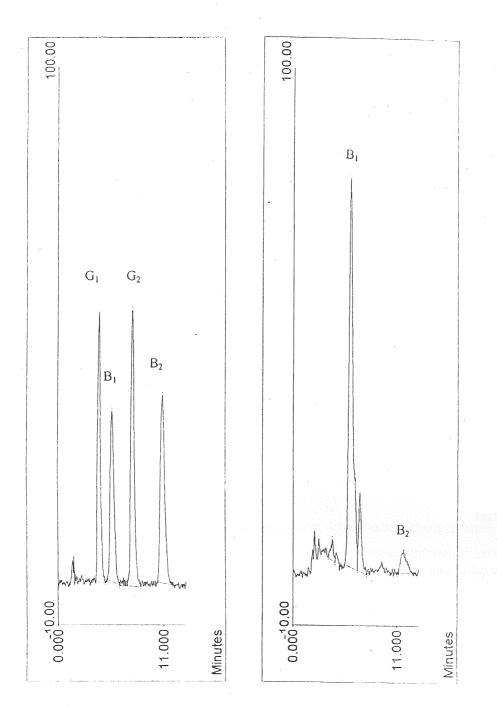


Figure 1. HPLC chromatograms: a) Aflatoxin standards (AF G_1 , B_1 , G_2 , B_2 ; 400 pg), b) Naturally contaminated dried red pepper extract (AF B_1 ;38.2 $\mu g.kg^{-1}$, B_2 ;2.3 $\mu g.kg^{-1}$)

Table 1. HPLC results of aflatoxins in red pepper and feedstuff specimens. Origin of specimens in Turkey are; UO: Unknown Origin, SE: Southeast, NW: Northwest, S: South, ND: Not Detected. (Results are in ppb).

	A 17-	Red pepper			Feedstuff
	AFs -	SE	S	UO	NW
AFs Positive Total	B_1	5/8	2/5	10/13	9/53
	B_2	2/8	0/5	3/13	3/53
	G_1	0/8	0/5	1/13	1/53
	G_2	0/8	0/5	0/13	0/53
Range	B ₁	3.1-56	2.7-10.3	0.6-38.2	5.1-59.9
	B_2	1.3-1.6	ND	0.4-2.3	15.3-48.7
	G_1	ND	ND	0.6	29.3
	G_2	ND	ND	ND	ND
Means of AFs positive	B ₁	22.94	6.50	16.94	28.81
	B_2	1.45	-	1.47	30.66
	G_1	-	-	0.6	29.30
	G_2	-	-	-	-

Özet

Bu çalışmanın amacı, Türkiye'de tüketilen kurutulmuş kırmızı biber ve yemlerde aflatoksin (AF) B₁, B₂, G₁ and G₂ 'yi araştırmaktır. AF analizinde yüksek basınçlı sıvı kromatografisi (HPLC) kullanıldı ve 1 ppb den yüksek sonuçlar ince tabaka kromatografisi (İTK) ile doğrulandı. HPLC'de B₁ ve G₁'in deteksiyon sınırı 0.75 ng ve B₂ ve G₂'nin deteksiyon sınırı 0.060 ng'dır. İTK da, AF lerin deteksiyon sınırı 1 ng dır. Piyasadan temin edilen toplam 26 kurutulmuş kırmızı biber ve 53 yem örneği analiz edildi. Türkiye'de baharatlarda AFB₁ için kabul edilen sınır değer 5 ppb ve tüm gıdalar için toplam AF düzeyi 10 ppb'dir. Bu çalışmada, AFB₁ 'i 5 ppb üzerinde içeren kurutulmuş kırmızı biber örneklerinin miktarı %42.3 ve toplam AF miktarını 10 ppb üzerinde içerenlerin miktarı %38.4 olarak

saptandı. Türkiye'de karma yemlerde toplam AF için kabul edilen sınır değer 50 ppb olup, örneklerin %7.5' u bu sınır değeri aşmıştır.

References

Adachi, Y., Hara, M., Kumazawa, H., Hirano, K., Ueno, I., Egawa, K. (1991). Detection of aflatoxin B₁ in imported food products into Japan by enzyme-linked immunosorbent assay and high performance liquid chromatography. *J.Vet.Med.Sci.* 53: 49-52.

Arim, R., Aguinaldo, A.R., Tanaka, T., Yoshizawa, T. (1999). Optimization and validation of a minicolumn method for determining aflatoxins in copra meal. *J.AOAC Int.* 82: 877-882.

Cavalheiro, A.C.L. (1983). Aflatoxin and aflatoxicosis. Zootecnica International 5: 41-43.

Ciegler, A. (1975). Mycotoxins: Occurrence chemistry, biological activity. *Lloydia*, 38: 21-35.

Çamlıbel, L. (1995). IGEME Research and Development Presidency, Agricultural Administration 1-23.

Dunne, C., Meaney, M., Smyth, M. (1993). Multimycotoxin detection and clean-up method for aflatoxins, ochratoxin and zearalenone in animal feed ingredients using high-performance liquid chromatography and gel permeation chromatography. *J. Chromatogr.*, 629: 229-235.

Ehrlich, K.C., Lee, L. (1984). Mycotoxins in grain dust: method for analysis of aflatoxins, ochratoxin A, zearalenone, vomitoxin, and secalonic acid D. *J.Assoc. Off. Anal. Chem.* 67: 963-967.

FAO (Food & Agriculture Organisation). (1997). Worldwide Regulations for Mycotoxin 1995. A compendium. FAO Food and Nutrition: Paper 64. (Rome: Food and Agriculture Organisation of the United Nations)

Garner, R.C., Whattam, M.M., Taylor, P.J.L., Stow, M.W. (1993). Analysis of United Kingdom purchased spices for aflatoxins using an immunoaffinity column clean-up procedure followed by high-performance liquid chromatographic analysis and post-column derivatisation with pyridinium bromide perbromide. *J. Chromatogr.*, 648: 485-490.

Harrison, J.C., Carvajal, M., Garner, R.C. (1993). Does aflatoxin exposure in the United Kingdom constitute a cancer risk?. *Environ. Health Persp.*, 99: 99-105.

Holcomb, M., Wilson, D.M., Trucksess, M.W., Thompson, H.C. (1992). Determination of aflatoxins in food products by chromatography. *J. Chromatogr.*, 624: 341-352.

Howell, M.V., Taylor, P.W. (1981). Determination of aflatoxins, ochratoxin A, and zearalenone in mixed feeds, with detection by thin layer chromatography or high performance liquid chromatography. *J.Assoc. Off. Anal. Chem.* 64: 1356-1363.

Ioannou-Kakouri, E., Aletrari, M., Christou, E., Hadjioannou-Ralli, A., Koliou, A., Akkelidou, D. (1999). Surveillance and control of aflatoxins B_1 , B_2 , G_1 , G_2 , and M_1 in foodstuffs in the republic of Cyprus: 1992-1996, *J.AOAC Int.* 82: 883-892.

Kichou, F., Walser, M.M. (1993). The natural occurrence of aflatoxin B_1 in Moroccan poultry feeds. *Vet. Hum. Toxicol*, 35:105-108.

Macdonald, S., Castle, L. (1996). A UK retail survey of aflatoxins in herbs and spices and their fate during cooking. *Food Addit. Contam.*, 13: 121-128.

Majerus, P., Zakaria, Z.Z. (1992). A rapid, sensitive and economic method for the detection, quantification and confirmation of aflatoxins. *Z Lebensm Unters Forsch*, 195: 316-319.

Nesheim, S., Trucksess, M.W., Page, S.W. (1999). Molar absorptivities of aflatoxins B_1 , B_2 , G_1 , and G_2 in acetonitrile, methanol, and toluene-acetonitrile (9+1) modification of AOAC official method 971.22): Collaborative study. *J.AOAC Int.* 82: 251-258.

Omurtag, G.Z. (2001). Determination of fumonisin B₁ and B₂ in corn and corn-based products by HPLC", *J. Food Prot.*, 64: 1072-1075.

Omurtag, G.Z., Atak, G., Yurdun, T., Ersoy, Ö. (1998). Aflatoxin analysis in dried red pepper samples by TLC and HPLC. *Acta Pharm. Turcica*, XXXX: 125-130.

Omurtag, G.Z., Yazıcıoğlu, D. (2000). Determination of T-2 toxin in grain and grain products by HPLC and TLC. J. Environ. Sci. Health, Part B, 35: 797-807.

Omurtag, G.Z., Yazıcıoğlu, D. (2001). Occurence of T-2 toxin in processed cereals and pulses in Turkey determined by HPLC and TLC, *Food Addit. Contam.*, 18, 844-849.

Özkazanç, A.N., Russel-Sin, H., Şanlı, Y., Kaya, S. (1992). The investigation of the pollution status arising from mycotoxins in the mixed feeds and feedstuffs produced in various region of Turkey. *J. Fac. Vet. Univ. Ankara*, 39, 268-290.

Paulsch, W.E., Sizoo, E.A., Van Egmond, H.P. (1988). Liquid chromatographic determination of aflatoxins in feedstuffs containing citrus pulp. *J. Assoc. Off. Anal. Chem.* 71: 957-961.

Scott, P.M. (1978). Mycotoxins in feeds and ingredients and their origin. *J. Food Prot.*, 41: 385-398.

T.C. Resmi Gazete (1990). 2 Mayıs 1990, No. 20506, 21.

T.C. Resmi Gazete (1997): 16 Kasım, Sayı No. 23172, 124.

Tabata, S., Kamimura, H., Ibe, A., Hashimoto, H., Iida, M., Tamura, Y., Nishima, T. (1993). Aflatoxin contamination in foods and feedstuffs in Tokyo: 1986-1990. *J.AOAC Int.* 76: 32-35.

Trucksess, M.W., Young, K., Donahue, K.F., Morris, D.K., Lewis, E. (1990). Comparison of two immunochemical methods with thin-layer chromatographic methods for determination of aflatoxins. *J. Assoc. Off. Anal. Chem.*, 73: 425-428.

WHO (1987). Supplement 1, Lyon, France, p.82

Accepted 24.10.2001