Nutritional Scrutiny, Chemical Profiling and Antioxidant Potentials of Crude Extracts of *Moringa oleifera* Lam. (Moringaceae) from Kasur, Pakistan

Muhammad Khalid SAEED¹, Naseem ZAHRA¹, Adil HUSSAIN^{1*}, Asma SAEED¹ Quratulain SYED¹

1 Food and Biotechnology Research Centre, Pakistan Council of Scientific and Industrial Research (PCSIR) Laboratories Complex, Ferozepur Road Lahore 54600 Punjab, Pakistan

ABSTRACT

Moringa oleifera L. is a miraculous plant due to the presence of essential nutrients, phyto-constituents and natural antioxidants. This study evaluated the aqueous and methanol extracts of *M. oleifera* leaves for various nutritional parameters, phytoconstituents and antioxidant activities. Results indicated varying contents of moisture (78.0 \pm 2.2, 7.3 \pm 0.7%), ash (0.8 \pm 0.1, 1.50 \pm 0.8%), fat (0.9 \pm 0.2, 1.8 \pm 0.9%), crude fiber (2.1±0.3, 20.5±1.7%), crude protein (7.1±0.8, 22.7±1.9%) carbohydrates (11.1±1.3, 46.2±2.1%) and energy kcal/100g (81.0±2.5, 292±4.2). Proteins, carbohydrates, hydroxyl-anthraquinone, tannins, alkaloids, saponins, flavonoids, terpenoids and saponins were present in Moringa leaves powder except phytosterol and fixed oil. In the methanol extract of Moringa fresh leaves, TPC recorded was 76.7±2.5 mg GAE/g and TFC was 24.6±0.40 mg QE/g while in the dried leaves powder, TPC and TFC were 86.2±1.8 mg GAE/g and 29.8±0.4 mg QE/g which were higher than the aqueous extracts. Antioxidant activity of Moringa dried leaves methanol extract with DPPH displayed maximum percentage inhibition $(92.5\pm3.2\%)$ than aqueous extract $(65.2\pm2.5\%)$ and BHT $(57.6\pm2.1\%)$ at 100μ g/ml. Same tendency was observed in the reducing power assay for Moringa dried leaves powder methanol extract Conclusively, M. oleifera possess a wealth of nutrients and bioactive compounds with potential antioxidant activity for extraordinary applications in the food and pharmaceutical industries.

*Corresponding Author: E-Mail: aadil.iiu07@gmail.com ORCIDS:

Muhammad Khalid SAEED: 0000-0003-0613-0896

Naseem ZAHRA: 0000-0002-3993-5079

Adil HUSSAIN: 0000-0002-8611-322X

Asma SAEED: 0009-0008-6370-0692

Quratulain SYED: 0009-0002-4412-6302

⁽Received 4 Feb 2023, Accepted 13 Apr 2023)

Keywords: *Moringa oleifera*, nutritional analysis, phyto-constituents, antioxidant activity, Pakistan

INTRODUCTION

Since the ancient time, plants are regarded as a significant source of medicine. According to the World Health Organization, up to 80% population of the world is still dependent on herbal preparations as medicine to treat different diseases¹. The Moringa oleifera Lam. is a famine resistant plant from the Moringaceae family². It is a plant of tropical forests whose all parts including the gum, seed, fruit, flowers, leaves, bark and roots are rich in proteins, vitamins and minerals like calcium, phosphorous, potassium, folic β -carotene, iron and acid. The fresh leaves or dried leaves powders of Moringa are employed for the development of food products to have improved nutritional quality and therapeutic effects³. In the ancient times, leaves of this plant were given to animals as feed⁴ and were used in human diet for better health. As its popularity rises, its various parts like roots, pods and seeds were found to be nutritious and medicinally significant. That's why; this plant is taken as a food ingredient, nutraceutical and medicine due to the presence of essential phytochemicals. It has been said that the phytochemicals in different parts of a particular plant makes it very significant and versatile medicinally5-7. Numerous phytochemicals have been extracted and reported from Moringa, including phenolics, flavonoids, tannins, alkaloids, saponins and glucosides etc. Polyphenolic compounds like flavonoids and phenolic acids are abundantly present in the dried leaves of Moringa^{9,10}.

Previously, phytochemicals like carbonic acid, 2-Isopropoxyethyl propionate (16.87%), 1,3-dioxolan-2-one, 4,5-dimethyl- (6.16%), 4H-pyran-4-one,2,3dihydro-3,5-dihydroxy-6-methyl- (8.98%), 1,3-dihydroxyacetone dimer (3.85%), 2-hydroxy-2-methyl- (3.14%), butyl 2-pentyl ester (20.64%), alpha-d-glucose (3.44%), azetidin-2-one 3,3-dimethyl-4-(1-aminoethyl)- (4.67%), tetra acetyl-dxylonic nitrile (5.03%) and butanedioic acid, were documented in *M. oleifera* leaves aqueous extracts. Compounds like 2-ethyl-2-(hydroxymethyl)- (21.19%), 1,3-propanediol, 2-methyl-, octyl ester (15.02%), propionic acid, n-ethyl-n-nitroso- (5.21%), ethanamine, 9,12,15-octadecatrienoic acid,(*Z*,*Z*,*Z*)- (5.00%), 4H-pyran-4-one,2,3dihydro-3,5-dihydroxy-6-methyl- (4.18%), benzeneacetonitrile, monomethyl malonate (2.56%), n-hexadecanoic acid (2.57%), 3-oeoxy-d-mannoic lactone (3.29%) were also recognized in the methanol extracts of *M. oleifera* leaves¹¹.

The phytochemicals from *Moringa* leaves and seeds have been reported with therapeutic potencies^{12,13} against cardiovascular diseases¹⁴, hypolipidemic disorders¹⁵, with biological activities including antispasmodic and diuretic¹⁶, antiobesity¹⁷, antiulcer¹⁸, antihypertensive^{19,20}, antitumor and apoptotic^{21,22}, hepatoprotective^{23,24}, antidiabetic²⁵⁻²⁷, antimicrobial²⁸, wound healing²⁹, analgesic³⁰, antipyretic³¹, antiasthmatic³², anti-inflammatory^{33,34}, antiurolithiatic³⁵ and antioxidant^{36,37} activities. *Moringa* plant have also protective effects against immune disorders³⁸ and neurodegenerative diseases including Parkinson's³⁹ and Alzheimer's⁴⁰.

As there is emergent interest in the assessment of nutritional and therapeutic efficacies of natural compounds from plants origin with their utilization in the development of products and drugs in the food and pharmaceutical industries, this study further extents the nutritional parameters, phytochemical profiles and antioxidant potentials of aqueous and methanol extracts of *M. oleifera* fresh leaves and dried leaves powder from the Kasur district of Pakistan.

METHODOLOGY

Moringa sample preparation

Fresh leaves of *M. oleifera* were acquired from the Kasur district of Pakistan and the collected specimens were identified by experts from the Government College University, Lahore, Pakistan. Primarily, the leaves were cleaned up with distilled water and placed in aluminum trays at 27°C for 5 days to shade dry. The dried leaf specimen was then grinded (Grinding mill, Germany) and the obtained powder was sieved with a 2 mm pore size siever and stored at -4°C for further experimentation⁴¹.

Nutritonal evaluation

For moisture analysis, *Moringa* fresh leaves and dried leaves powder were placed in oven at 105 to 110°C to a constant weight. Nutritional parameters like, crude lipid, crude protein, crude fiber and total ash content were estimated by adapting AOAC standard methods⁴². Estimation of total carbohydrates was done using the method described in literature^{43,44}. The values of energy in Kcal/100 g were estimated by multiplying obtained values of carbohydrates, lipids and proteins by factors of 4 and 9 and the sum obtained was presented in kilocalories⁴⁵.

Solvent extraction and phytochemical analysis

A 1% (w/v) stock concentration of extract was obtained from fresh and dried leaves powder of *Moringa* using methanol and water as extraction solvents. Following standard procedures given by Harborne⁴⁶ and Kokate⁴⁷, the *Moringa* extracts were tested with positive and negative controls for qualitative testing of phytochemicals like amino acids, alkaloids, triterpenoids, tannins, phytosterols, anthroquinone, flavonoids, cardiac glycosides, carbohydrates, glycosides, saponins, fixed oils/fats and proteins.

Estimation of TPC and TFC

The total phenolic content (TPC) of *Moringa* fresh leaves and dried leaves powder were determined quantitatively by folin-ciocalteau reagent method at 760 nm⁴⁸ with some modifications⁴⁹ and the obtained results were presented in mg GAE/100g. For the total flavonoid content (TFC) estimation in *M. oleifera* fresh leaves and dried leaves powder, the aluminium chloride colorimetric method was used⁵⁰ with minor modifications⁵¹. Quercetin was used as a standard compound for flavonoids quantification and the obtained values were presented as mg QE/100g.

Antioxidant potentials of Moringa

The free radical scavenging potency of *M. oleifera* fresh leaves and dried leaves powder was assessed using DPPH method⁵² with some modifications^{53,54} where the antioxidants minimize the free radicals absorbing light at 517 nm. The ability of aqueous and methanol extracts of *Moringa* fresh leaves and dried leaves powder to reduce iron (III) to iron (II) (total reducing power) was evaluated following Oyaizu⁵⁵ with slight modification^{53,54} and compared to a strong reducing agent BHT. The absorbance (700 nm) of the samples was plotted against each concentration taken.

Statistical analysis

Data was analyzed statistically and standard deviation (SD \pm) was estimated in the Microsoft excel program. Differences at p < 0.05 were considered significant⁵⁶.

RESULTS and DISCUSSION

Nutritional evaluation of M. oleifera

Nutritional analysis of *M. oleifera* fresh leaves and dried leaves powder performed using the proximate analysis is critical in determining the nutritional quality of *Moringa*. The results of nutritional parameters assessed in *M. oleifera* leaves are given in Table 1 where the fresh leaves and dried leaves powder exhibited varying contents of moisture (78.0 ± 2.2 , $7.3 \pm 0.7\%$), ash (0.9 ± 0.1 , $6.50 \pm 0.8\%$), fat (0.8 ± 0.2 , $1.8 \pm 0.9\%$), crude fiber (2.1 ± 0.3 , $20.5 \pm 1.7\%$), crude protein (7.1 ± 0.8 , $22.7 \pm 1.9\%$), carbohydrates (11.1 ± 1.3 , $46.2 \pm 2.1\%$). The total energy estimated in *Moringa* fresh leaves and dried leaves powder were 81.0 ± 2.5 , 272 ± 3.2 Kcal/100g.

These results regarding the moisture and ash contents recorded in *Moringa* dried leaf powder (7.3%, 6.5%) are in line with the data reported in previous studies on *Moringa* where mean contents of moisture recorded were between

5 -10% and ash between 6-11%^{57,58}. In other investigations, elevated levels of moisture and slightly low ash contents were reported in *Moringa* on the basis of dryness of the original sample^{59,60}. The ash of *Moringa* may possess inorganic minerals that are necessary for growth and development. It has been found that *Moringa* possess more Ca and iron as compared to spinach, that's why; the leaves powder of *Moringa* could be utilized as a substitute for iron medications for the treatment for anemia. It has been also found that *Moringa* leaves have around 25.5–31.03 mg of zinc/kg⁶¹.

The mean protein contents of fresh leaves and dried leaves powder of *Moringa* were 7.1±0.8 and 22.7 ±1.9% respectively. These results regarding the protein contents are in accordance with the data reported by Valdez-Solana *et al.*⁶² and Fejér *et al.*⁶³. However, Moyo *et al.*⁶⁴, Castillo-López *et al.*⁶⁵ and Fokwen *et al.*⁶⁶ reported slightly lower (27.2%) protein contents in *Moringa*.

Other nutritional parameters assessed in fresh leaves and dried leaves powder of *Moringa* include fat (0.9±0.2, 1.8±0.9%), crude fiber (2.1±0.3, 20.5±1.7%), crude protein (7.1±0.8, 22.7±1.9%) and carbohydrates (11.1±1.3, 46.2±2.1%). These outcomes are consistent with the results of earlier inquiries of Yameogo *et al.*⁶⁷, Witt⁶⁸ and Isitua *et al.*⁶⁹. The nutrient variation in *Moringa* could be due to the seasonal fluctuations such as climate, location, geography and some other environmental factors⁷⁰. The physicochemical analysis confirmed that *M. oleifera* is a miraculous plant and its leaves are a high quality food source making itself a candidate plant to be utilized directly in human diet or in the improvement of balanced diets in animal nutrition⁷¹. In spite of this difference, proximate analysis displayed that the leaves of *M. oleifera* remained good sources of fats, fiber, carbohydrate and proteins which are the primary sources of energy and are used in food and medicinal products like bread, biscuits, drinks, cookies, soups and medicinally coated capsules.

Percentage values			
Moringa leaves (Fresh)	<i>Moringa</i> dried leaves (Powder)		
78.0±2.2	7.3±0.7		
0.9±0.1	6.5±0.8		
0.8±0.1	1.8±0.9		
2.1±0.3	20.5±1.7		
7.1±0.8	22.7±1.9		
11.1±1.3	41.2±2.1		
81±2.5	272±3.2		
	Moringa leaves (Fresh) 78.0±2.2 0.9±0.1 0.8±0.1 2.1±0.3 7.1±0.8 11.1±1.3		

Table 1. Results of nutritional parameters of *M. oleifera* fresh leaves and dried leaves powder

Values are the mean (\pm SD) of three readings presented as (%) g/100 g

Phytochemical analysis of M. oleifera

Basic phytochemical analysis for confirming the presence of major phytocompounds is critical because many drugs' active principles are secondary metabolites present in plants. Findings showed that the aqueous and methanol extracts of *Moringa* leaves possess many phytochemical groups including tannins, alkaloids, saponins, flavonoids, protein, terpenoids, hydroxyl anthraquinone and carbohydrates, but no phytosterol or fixed oil was found in both the extracts (Table 2). These outcomes are in accordance with the results of Vergara-Jimenez et al.72, Ayoade et al.73, and Sudha et al.74. The utilization of methanol and water as extraction solvents indicated the presence of assorted active principles with solvents selective solubility with different polarities used in succession, inferring the solvents significance as a promising factor⁷⁵. Furthermore, the evidence points to the significance of a particular test as a key factor in endorsing phytochemicals presence. Since *M. oleifera* leaves are rich in amino acids and carbohydrates, it is proposed that these nutrients are very beneficial and may be utilized as a growth promoters and nutritional supplements^{76,77}. Phytochemical data of *M. oleifera* may be used to produce lead compounds in the quest for innovative herbal medications^{78,79}.

Phytochemicals	Tests/ Experiments	Blank	Control	Aqueous extract	Methanol extract
Tannin	Ferric chloride	-	+++	+	++
Alkaloids	Dragendorff's test	-	++	+	++
Saponins	Foam test	- ++		-	+
Triterpenoids	Salkowski	-	+++	++	+++
Phytosterols	Liebermann- Burchard	-	+++	-	-
Cardiac glycoside	Keller killani	-	+++	-	+
Flavonoids	Lead acetate	-	+++	++	+++
Hydroxyanthraquinone	Potassium hydroxide	-	+++	++	+
Amino acid	Millon's test	-	++	+	++
Fixed oils and fats	Copper sulphate	-	++	-	-
Carbohydrates	Molisch's test	-	+++	++	+++
Proteins	Biuret test	-	++	+	++

Table 2. Phytochemical profile of aqueous and methanol extracts of *M. oleifera* fresh leaves

++++ (Very high), +++ (High), ++ (Moderate), + (Low), - (Nil), Positive controls and blank (water)

Total phenolic content (TPC) in M. oleifera

Phenolic compounds are much prevalent in plants with potential antioxidant activities as they generate hydrogen ions that form stable intermediate radicals⁸⁰. This study assessed the TPC with folin-ciocalteu method where the *Moringa* fresh leaves aqueous extract displayed lower (60.2±1.3 mg GAE/g) and the methanol extract showed higher TPC content (76.5±1.7 mg GAE/g). While, the aqueous extract of dried leaves powder contains 68.3±1.5 mg GAE/g and methanol extract has TPC of 86.2±1.8 mg GAE/g (Figure 1, Table 3). The TPC reported here are faintly lower as compared to the TPC content (9535.3±57.74 mg/100g) reported by Ilyas *et al.*⁸¹ whereas; slightly higher values of TPC were described in *Moringa* leaves by Abdulkadir *et al.*⁸². The variations in TPC might be associated with the difference in polyphenolics extraction methods or solvents polarity and also the plants geographical distribution. Previous data shows that the *Moringa* leaves are rich in valued compounds like vitamin, protein, calcium, iron and antioxidants including ascorbic acid, carotenoids, phenols and flavonoids⁸³. Due to the polyphenols and other antioxidants, many researchers have claimed that the methanol extract of *M. oleifera* leaves exhibit a strong antioxidant action⁸⁴. Many studies have also proposed that the *Moringa* leaves possess anti-diabetic, anti-inflammatory, anti-epileptic, anti-hypertensive, and antitumor activities and these are associated with the phenolic compounds present in the plant. Further, the phenolic compounds have strong antioxidant activities against tissue impairments instigated by the free radicals^{85,86}.

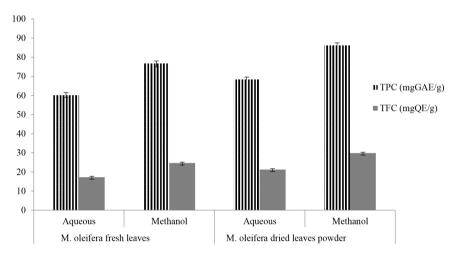


Figure 1. Total phenolic and flavonoid contents in methanol and aqueous extracts of *M. oleifera* fresh leaves and dried leaves powder

Extracts	M. oleifera fresh leaves		<i>M. oleifera</i> dried leaves powder	
	TPC (mgGAE/g)	TFC (mgQE/g)	TPC (mgGAE/g)	TFC (mgQE/g)
Aqueous (H ₂ O)	60.2±1.3	17.2±0.2	68.3±1.5	21.2±0.3
Methanol (MeOH)	76.7±1.7	24.6±0.5	86.2±1.8	29.8±0.4

Table 3. Total phenolic and flavonoid contents in *M. oleifera* fresh leaves and dried leaves powder

Data are represented as mean $(\pm SD)$ of three readings

Total flavonoid content (TFC) in M. oleifera

It was found that the methanol extract of *Moringa* dried leaves powder has greater concentration of flavonoids (29.6±0.4 mg QE/g) than its aqueous extract (21.2±0.3 mg QE/g) (Figure 1, Table 3). Flavonoids were confirmed in both the aqueous and methanol extracts of *Moringa* fresh leaves and dried leaves powder. According to Lin *et al.*⁸⁷, flavonoids are polyphenolic compounds mostly found in dried leaves of *Moringa*. They are secondary metabolites which are most prevalent phytochemical group. According to Masood *et al.*⁸⁸, plant's antioxidant capacity is correlated with its level of TPC and TFC. Flavonoids have positive effects on the human body and provides protection against various diseases⁴⁹. The leaves of *M. oleifera* contain variety of flavonoids; however the most prevalent flavonoids with significant pharmacological action are quercetin, apigenin, kaempferol and isorhamnetin⁹⁰. Flavonoids also possess potential anti-microbial, anti-inflammatory, antioxidant, anti-allergic potentials and other significant biological activities^{91,92}.

Antioxidant activity of M. oleifera extracts

The free radical scavenging potentials of plant extracts improves with the increase in extract concentration. This pattern of radical scavenging was observed in the present inquiry where the percentage inhibitions recorded in methanol extract of *Moringa* dried leaves powder were 35.5, 50.2, 65.9, 78.1, and 92.5% at 20, 40, 60, 80, and 100 μ g/ml concentrations , while the percentage inhibitions of the aqueous extract at same concentrations were 25.4, 34.1, 45.6, 55.3, and 65.2 respectively. Similarly, the percentage inhibitions of the standard synthetic antioxidant BHT at same concentrations recorded were 21.5, 30.2, 40.7, 48.3, and 57.6% (Figure 2). The aqueous and methanol extracts of *Moringa* dried leaves powder showed the similar patterns of DPPH scavenging activities which are higher than BHT.

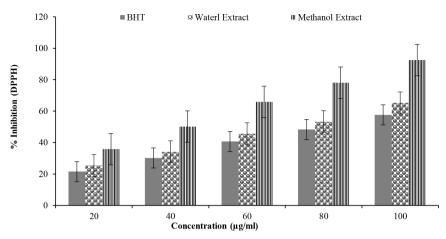


Figure 2. Percentage inhibition (DPPH) of aqueous and methanol extracts of *M. oleifera* dried leaves powder and BHT

The phytochemicals bear antioxidant properties due to the ability to prevent production free radicals, or scavenging free radicals in the body or chelating/ reducing the content of transition metal^{93,94}. An essential antioxidant method is considered to be the inhibition of the chain start phase by scavenging different reactive species like the free radicals^{53,54,95,96}.

ROS (reactive oxygen species) are continuously generated in animals due to some environmental factors encountered in daily life⁹⁷. In such conditions, antioxidants are generated by the body cells to maintain the body's equilibrium with free radicals. Oxidative stress describes any imbalance brought on by a variety of diseases in the regular physiological system of the body. According to Karim *et al.*⁹⁸ at the harsh level, the oxidative stage transforms cellular damage into different chronic diseases. It has been found that the antioxidants have a good effect on these types of chronic diseases by preventing the initiation of any damage⁹⁹⁻¹⁰¹.

As per the findings of this study, the methanol extract of *Moringa* dried leaves powder displayed higher percentage of DPPH inhibition than the aqueous extract and BHT which are in accordance with previous investigations of Kumar *et al.*¹⁰², Almaghrabi *et al.*¹⁰³ and Landazuri *et al.*¹⁰⁴.

The increased DPPH scavenging activity of methanol extracts of *Moringa* reported here might be associated with the presence of total phenolic and flavonoid contents. By providing hydrogen atoms to DPPH, these hydroxyl phenolic compounds can remove it from the environment. In order to evaluate the antioxidant potentials of herbal extracts, the DPPH scavenging method is now frequently used¹⁰⁵⁻¹⁰⁷ which is highly accurate, sensitive, and quick, depends on the transformation of unstable purple DPPH molecules into yellowish DPPH molecules in the presence of antioxidants^{108,109}.

The reducing power of *Moringa* dried leaves powder extract was assessed and compared with standard reference BHT on ferric to ferrous reduction in the presence of Fe (II) - stabilising ligand (Figure 3). The reducing power might be attributed to the hydrogen donating capacity, which is commonly related with the presence of reductants¹¹⁰. The extracts can convert [Fe (CN)₆]₃- to [Fe(CN)₆]₄-, which then interacts with Fe³⁺ to form Fe₄[Fe(CN)₆]₃ which is a Prussian blue coloured complex¹¹¹. Our findings revealed that these extracts have a degree of hydrogen donation ability which varies with concentration.

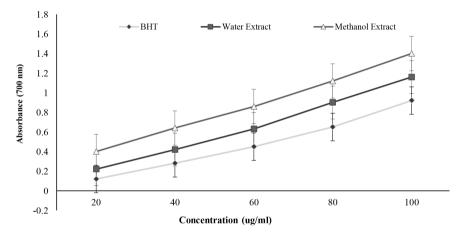


Figure 3. Reducing power of aqueous and methanol extracts of *M. oleifera* dried leaves powder and BHT

The reducing power increased with concentration and these capabilities were superior to those of standard synthetic antioxidant BHT. *Moringa* dried leaves powder extracted with methanol was found to be the most powerful reducing agent, followed by the aqueous extract and BHT. These findings form a clear relationship between the reduction efficacy and antioxidant capability of *Moringa* extracts with high phenolic content as the phenols with a greater number of hydrolysable groups (OH groups) connected to the ring are potent reducing agents (proton donors), resulting in the termination of free radical chain reactions¹¹².

Considering its myriad benefits, *Moringa* really does seem to be a Marvel plant. As such, this plant must be adapted as a high-quality, inexpensive memento from nature. In addition to its remarkable health benefits, this study found that the crude methanol and aqueous extracts of *M. oleifera* leaves possess essential nutrients, phytochemicals and natural antioxidants that could be significant for industrial and medicinal purposes. To have improved nutrition, the extracts of *Moringa* dried leaves powder should be used for better effects. To fully investigate and utilize the wonders of the *Moringa* tree, more robust research and product development strategies are required. The moment has come to investigate its route for food usage, standardize and commercialize technology for producing value-added and highly nutritious products.

CONFLICT OF INTEREST STATEMENT

Nothing to declare.

AUTHOR CONTRIBUTIONS

Design: Muhammad Khalid SAEED

Acquisition of data: Muhammad Khalid SAEED

Analysis of data: Muhammad Khalid SAEED

Drafting of the manuscript: Muhammad Khalid SAEED and Adil HUSSAIN

Critical revision of the manuscript: Muhammad Khalid SAEED and Adil HUS-SAIN

Statistically analysis: Muhammad Khalid SAEED

Technical and Financial Support: Muhammad Khalid SAEED, Naseem ZAH-RA (these authors contributed equally)

Supervision: Muhammad Khalid SAEED, Asma SAEED and Quratulain SYED (these authors contributed equally)

Other (Specify): NA

FUNDING SOURCES

None.

ACKNOWLEDGMENTS

The author's extent gratefulness to the anonymous reviewers for their appreciated suggestions to improve the quality of this manuscript.

REFERENCES

1. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. Front Pharmacol, 2014;4:1–10. https://doi.org/10.3389/fphar.2013.00177

2. Dhakad AK, Ikram M, Sharma S, Khan S, Pandey VV, Singh A. Biological, nutritional, and therapeutic significance of *Moringa oleifera* Lam. Phyther Res, 2019;33(11):2870–2903. https://doi.org/10.1002/ptr.6475

3. Islam Z, Islam SMR, Hossen F, Mahtab-Ul-Islam K, Hasan MR, Karim R. *Moringa oleifera* is a prominent source of nutrients with potential healthbenefits. Int J Food Sci, 2021;6627265. https://doi.org/10.1155/2021/6627265

4. Sun J, Zeng B, Chen Z, Yan S, Huang W, Sun B, He Q, Chen X, Chen T, Jiang Q, Xi Q, Zhang Y. Characterization of faecal microbial communities of dairy cows fed diets containing ensiled *Moringa oleifera* fodder. Scient Rep, 2017;7:41403. https://doi.org/10.1038/srep41403

5. Jan H, Usman H, Shah M, Zaman G, Mushtaq S, Drouet S, Hano C, Abbasi BH. Phytochemical analysis and versatile in vitro evaluation of antimicrobial, cytotoxic and enzyme inhibition potential of different extracts of traditionally used *Aquilegia pubiflora* Wall. Ex Royle. BMC Complement Med Ther, 2021; 21(1):165. https://doi.org/10.1186/s12906-021-03333-y

6. Liang L, Wang C, Li S, Chu X, Sun K. Nutritional compositions of Indian *Moringa oleifera* seed and antioxidant activity of its polypeptides. Food Sci Nut, 2019;7(5):1754–1760. https://doi.org/10.1002/fsn3.1015

7. Hussain A, Hayat MQ, Sahreen S, Bokhari SAI. Pharmacological promises of genus *Artemisia* (Asteraceae): a review. Proc Pak Acad Sci: B Life Env Sci, 2017;54:265–287.

8. Abalaka ME, Daniyan SY, Oyeleke SB, Adeyemo SO. The antibacterial evaluation of *Moringa oleifera* leaf extracts on selected bacterial pathogens. J Microbiol Res, 2012;2(2):1–4. https://doi.org/10.5923/j.microbiology.20120202.01

9. Hassan MA, Xu T, Tian Y, Zhong Y, Ali FAZ, Yang X, Lu B. Health benefits and phenolic compounds of *Moringa oleifera* leaves: A comprehensive review. Phytomedicine, 2021;93:153771. https://doi.org/10.1016/j.phymed.2021.153771

10. Makita C, Chimuka L, Steenkamp P, Cukrowska E, Madala E. Comparative analyses of flavonoid content in *Moringa oleifera* and *Moringa ovalifolia* with the aid of UHPLC-qTOF-MS fingerprinting. South Afr J Bot, 2016;105:116–22. https://doi.org/10.1016/j.sajb.2015.12.007

11. Bhalla N, Ingle N, Patri SV, Haranath D. Phytochemical analysis of *Moringa oleifera* leaves extracts by GC-MS and free radical scavenging potency for industrial applications. Saudi J Biol Sci, 2021;28: 6915–6928. https://doi.org/10.1016/j.sjbs.2021.07.0751

12. Demiray S, Pintado ME, Castro PML. Evaluation of phenolic profiles and antioxidant activities of Turkish medicinal plant: *Tilia argentea*, *Crataegi folium* leaves and *Polygonum bistorta* roots. World Acad Sci, Engin Technol, 2009;54:312–317.

13. Patil S, Mohite BV, Marathe KR, Salunkhe NS, Marathe V, Patil VS. Moringa Tree, Gift of Nature: a Review on Nutritional and Industrial Potential. Curr Pharmacol Rep, 2022;8:262–280. https://doi.org/10.1007/s40495-022-00288-7

14. Syed Muhammad AS, Muhammad A, Muhammad R, Naveed M, Ghulam R. Cardioprotective potential of plant-derived molecules: A scientific and medicinal approach. Dose-Response, 2019;1-14. https://doi.org/10.1177%2F1559325819852243

15. Jain PG, Patil SD, Haswani NG, Girase MV, Surana SJ. Hypolipidemic activity of Moringa

oleifera Lam., *Moringaceae*, on high fat diet induced hyperlipidemia in albino rats. Braz J Pharmacog, 2010; 20(6):969–73. https://doi.org/10.1590/S0102-695X2010005000038

16. Caceres A, Saravia A, Rizzo S, Zabala L, Leon ED, Nave F. Pharmacologic properties of *Moringa oleifera*: 2: Screening for antispasmodic, anti-inflammatory and diuretic activity. J Ethnopharmacol, 1992;36:233–237. https://doi.org/10.1016/0378-8741(92)90049-w

17. Bais S, Singh GS, Sharma R. Antiobesity and hypolipidemic activity of *Moringa oleifera* leaves against high fat diet-induced obesity in rats. Adv Biol, 2014;1–9. https://doi. org/10.1155/2014/162914

18. Choudhary MK, Bodakhe SH, Gupta SK. Assessment of the antiulcer potential of *Moringa oleifera* root-bark extract in rats. J Acupunct Merid Stud, 2013;6:214–220. https://doi. org/10.1016/j.jams.2013.07.003

19. Dangi SY, Jolly CI, Narayana S. Antihypertensive activity of the total alkaloids from the leaves of *Moringa oleifera*. Pharml Biol, 2002;40:144–148. https://doi.org/10.1076/phbi.40.2.144.5847

20. Randriamboavonjy JI, Rio M, Pacaud P, Loirand G, Tesse A. *Moringa oleifera* seeds attenuate vascular oxidative and nitrosative stresses in spontaneously hypertensive rats. Oxid Med Cell Longev. 2017; 4129459. https://doi.org/10.1155/2017/4129459

21. Adebayo IA, Arsad H, Kamal NM and Samian MR. The hexane fraction of the *Moringa oleifera* Lam seed extract induces apoptosis, causes cell cycle arrest, and modulates expression of HSP60, NPM, PGK1, RCN1, and PDIA1 in MCF7 cells. South Afr J Bot, 2019;1–9. https://doi.org/10.1016/j.sajb.2019.09.001

22. Shousha WG, Aboulthana WM, Salama AH, Saleh MH, Essawy EA. Evaluation of the biological activity of *Moringa oleifera* leaves extract after incorporating silver nanoparticles, *in-vitro* study. Bull Nat Res Cent, 2019;43:212. https://doi.org/10.1186/s42269-019-0221-8

23. Oyagbemi AA, Omobowale TO, Azeez IO, Abiola JO, Adedokun RA, Nottidge HO. Toxicological evaluations of methanolic extract of *Moringa oleifera* Leaves in liver and kidney of male Wistar rats. J Basic Clin Physiol Pharmacol, 2013;24:307–312. https://doi.org/10.1515/ jbcpp-2012-0061

24. Almatrafi MM, Vergara-Jimenez M, Murillo AG, Norris GH, Blesso CN, Fernandez ML. *Moringa* leaves prevent hepatic lipid accumulation and inflammation in guinea pigs by reducing the expression of genes involved in lipid metabolism. Int J Mol Sci, 2017;18:1330. https://doi.org/10.3390/ijms18071330

25. Divi SM, Bellamkonda R, Dasireddy SK. Evaluation of antidiabetic and antihyperlipedemic potential of aqueous extract of *Moringa oleifera* in fructose fed insulin resistant and STZ induced diabetic wistar rats: a comparative study. Asian J Pharm Clin Res, 2012;5(2012): 67–72.

26. Mbikay M. Therapeutic potential of *Moringa oleifera* leaves in chronic hyperglycemia and dyslipidemia: a review. Front Pharmacol, 2012;3:24. https://doi.org/10.3389/ fphar.2012.00024

27. Bienvenu T, Chougourou DC and Todohoue CM. Anti-hyperglycaemic and lipid profile regulatory properties of *Moringa oleifera* in subjects at early stages of type 2 diabetes mellitus. EMJ Diabet, 2016;4(1):99–105. https://doi.org/10.33590/emjdiabet/10310563

28. Abdallah R, Mostafa NY, Kirrella GAK, Gaballah I, Imre K, Morar A, Herman V, Sallam KI, Elshebrawy HA. Antimicrobial effect of *Moringa oleifera* leaves extract on foodborne pathogens in ground beef. Foods. 2023;12(4):766. https://doi.org/10.3390/foods12040766

29. Al-Ghanayem AA, Alhussaini MS, Asad M, Joseph B. *Moringa oleifera* Leaf Extract Promotes Healing of Infected Wounds in Diabetic Rats: Evidence of Antimicrobial, Antioxidant and Proliferative Properties. Pharmaceuticals (Basel), 2022;15(5):528. https://doi. org/10.3390/ph15050528

30. Martínez-González CL, Martínez L, Martínez-Ortiz EJ, González-Trujano ME, Déciga-Campos M, Ventura-Martínez R, Díaz-Reval I. *Moringa oleifera*, a species with potential analgesic and anti-inflammatory activities. Biomed Pharmacother, 2017;87:482–488. https://doi.org/10.1016/j.biopha.2016.12.107

31. Olaniran O, Adetuyi FC, Omoya FO, Odediran SA, Hassan-olajokun RE, Awoyeni EA, Odetoyin BW, Adesina A, Awe A, Bejide RA, Akinyemi LO, Oyetoke OO, Afolayan DO. Antiplasmodial, antipyretic, haematological and histological effects of the leaf extracts of *Moringa oleifera* in *Plasmodium berghei* berghei infected mice. J Adv Med Med Res, 2019;29(4):1–13. https://doi.org/10.9734/jammr/2019/v29i430083

32. Palupi DA, Prasetyowati TW, Murtiningsih D, Mahdiyah D. Antiasthma activities of *Moringa oleifera Lam.* leaves extract on the eosinophil count and mast cells in BALB/c mice. Borneo J Pharm [Internet]. 2021; 4(3):171-7. https://doi.org/10.33084/bjop.v4i3.1916

33. Zheng L, Lu X, Yang S, Zou Y, Zeng F, Xiong S, Cao Y, Zhou W. The anti-inflammatory activity of GABA-enriched *Moringa oleifera* leaves produced by fermentation with *Lactobacillus plantarum* LK-1. Front Nutr, 2023;10:1093036. https://doi.org/10.3389/ fnut.2023.1093036

34. Waterman C, Cheng DM, Rojas-Silva P, Poulev A, Dreifus J, Lila MA, Raskin I. Stable, water extractable isothiocyanates from *Moringa oleifera* leaves attenuate inflammation *in vitro*. Phytochemistry, 2014;103: 114-122. https://doi.org/10.1016/j.phytochem.2014.03.028

35. Karadi RV, Palkar MB, Gaviraj EN, Gadge NB, Mannur VS, Alagawadi KR. Antiurolithiatic property of *Moringa oleifera* root bark. Pharm Biol, 2008;46(12):861-865. https://doi.org/10.1080/13880200802367189

36. Luqman S, Srivastava S, Kumar R, Maurya AK, Chanda D. Experimental assessment of *Moringa oleifera* leaf and fruit for its antistress, antioxidant and scavenging potential using *in-vitro* and *in-vivo* assays. Evid Based Compl Alt Med, 2012;1–12. https://doi. org/10.1155/2012/519084

37. Peñalver R, Martínez-Zamora L, Lorenzo JM, Ros G, Nieto G. Nutritional and sntioxidant properties of *Moringa oleifera* leaves in functional foods. Foods, 2022;11(8):1107. https://doi.org/10.3390/foods11081107

38. Xiao X, Wang J, Meng C, Liang W, Wang T, Zhou B, Wang Y, Luo X, Gao L, Zhang L. *Moringa oleifera* Lam and its therapeutic effects in immune disorders. Front Pharmacol, 2020;11:566783. https://doi.org/10.3389/fphar.2020.566783

39. Giacoipo S, Rajan TS, De Nicola GR, Iori R, Rollin P, Bramanti P, Mazzon E. The isothiocyanate isolated from *Moringa oleifera* shows potent anti-inflammatory activity in the treatment of murine subacute Parkinson's disease. Rejuven Res, 2017;20:50–63. https://doi.org/10.1089/rej.2016.1828

40. Mahaman YAR, Huang F, Wu M, Wang Y, Wei Z, Bao J, Salissou MTM, Ke D, Wang Q, Liu R, Wang JZ, Zhang B, Chen D, Wang X. *Moringa oleifera* alleviates homocysteineinduced Alzheimer's disease-like pathology and cognitive impairments. J Alzheimers Dis, 2018;63(3):1141–1159. https://doi.org/10.3233/JAD-180091

41. Atawodi SE, Atawodi JC, Idakwo GA, Pfundstein B, Haubner R, Wurtele G, Bartsch H, Owen RW. Evaluation of the polyphenol content and antioxidant properties of met-

hanol extracts of the leaves, stem, and root barks of *Moringa oleifera* Lam. J Med Food, 2010;13(3):710–716. https://doi.org/10.1089/jmf.2009.0057

42. AOAC. Official methods of analysis, 21st ed. Gaithersburg, Maryland, U.S.A.: Association of Official Analytical Chemists, 2016.

43. Muller HG, Tobin G. Nutrition and food processing, Croom Helm, London, 1980.

44. Plummer DT. An introduction to practical biochemistry, 179 Third edition, 1990.

45. Imran M, Khan H, Hassan SS, Khan R. Physicochemical characteristics of various milk samples available in Pakistan. J Zhejiang Uni: Sci, 2008;B9(7):546–551. https://doi. org/10.1631/jzus.B0820052

46. Harborne JB. Textbook of phytochemical methods. A guide to modern techniques of plant analysis. 5th Edition, Chapman and Hall Ltd, London, 1998;21–72.

47. Kokate CK. A text book of practical pharmacognosy. 5th Edition, Vallabh Prakashan New Delhi, 2005;107–111.

48. Singleton VL, Rossi JA. 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. Am J Enol Viticul, 1965;16:144–158. https://doi. org/10.5344/ajev.1965.16.3.144

49. Saeed MK, Nisa A, Ahmad I, Hina S, Zahra N, Kalim I, Masood S, Syed Q. Physico-chemical analysis, total polyphenolic content and antioxidant capacity of yellow dye extracted from *Curcuma Longa*. Pak J Scient Ind Res, Ser. B: Biol Sci, 2021;64B(1):25–29. https://doi. org/10.52763/PJSIR.BIOL.SCI.64.1.2021.25.29

50. Dewanto V, Wu X, Adom KK, Liu RH. Thermal processing enhances the nutritional value of tomatoes by increasing total antioxidant activity. J Agric Food Chem, 2002;50:3010–3014. https://doi.org/10.1021/jf0115589

51. Saeed MK, Khan MN, Ahmad I, Hussain N, Ali S, Deng Y, Dai R. Isolation identification and antioxidant potential of major flavonoids from ethyl acetate fraction of *Torreya grandis*. Asian J Chem, 2013; 25(5):2459–2464. https://doi.org/10.14233/ajchem.2013.13401

52. Brand-Williams W, Cuvelier ME, Berset CLWT. 1995. Use of a free radical method to evaluate antioxidant activity. LWT-Food Sci Technol, 1995;28(1):25–30. https://doi. org/10.1016/S0023-6438(95)80008-5

53. Saeed MK, Zahra N, Abidi SHI, Syed Q. Phytochemical screening and DPPH free radical scavenging activity of *Aloe vera* (*Aloe barbadensis* Miller) powder. Int J Food Sci Agri, 2022a;6(3):301–308.

54. Saeed MK, Zahra N, Ahmad I, Shahzad K, Ashraf M, Abidi SHI. Syed Q, Arif K. Evaluation of nutritional parameters and antioxidant study of Musk melon powder from Lahore, Pakistan. Green Rep, 2022b;1(7): 30–33.

55. Oyaizu M. Studies on products of browning reactions: antioxidative activities of products of browning reaction prepared from glucosamine. Japan J Nut, 1986;44:307–315. https://doi.org/10.5264/eiyogakuzashi.44.307

56. Olawuyi JF. Biostatistics: A foundation course in health sciences. 1st Edition. University College Hospital, Published by Tunji Alabi Printing Co. Total Garden, Ibadan, Nigeria, 1996;1–221.

57. Rajput H, Prasad SGM, Srivastav P, Singh N, Suraj L, Chandra R. Chemical and phytochemical properties of fresh and dried *Moringa oleifera* (PKM-1) leaf powder. Chem Sci Rev Lett, 2017;6: 1004–1009.

58. Olusanya RN, Kolanisi U, van Onselen A, Ngobese NZ, Siwela M. Nutritional composition and consumer acceptability of *Moringa oleifera* leaf powder (MOLP)-supplemented mahewu. South Afr J Bot, 2020;129:175–180.

59. Amabye TG, Gebrehiwot K. Chemical compositions and nutritional value of *Moringa oleifera* available in the market of Mekelle. J Food Nut Sci, 2015;3:187. https://doi. org/10.11648/j.jfns.20150305.14

60. Umerah NN, Asouzu AI, Okoye JI. Effect of processing on the nutritional composition of *Moringa olifera* leaves and seeds. Eur J Nut Food Saf, 2019;11:124–135. https://doi. org/10.9734/ejnfs/2019/v11i330155

61. Barminas JT, Charles M, Emmanuel D. Mineral composition of non-conventional leafy vegetables. Plant Foods Hum Nutr, 1998;53(1):29–36. https://doi.org/10.1023/a:1008084007189

62. Valdez-Solana MA, Mejía-García VY, Téllez-Valencia A, García-Arenas G, Salas-Pacheco J, Alba-Romero JJ, Sierra-Campos E. Nutritional content and elemental and phytochemical analyses of *Moringa oleifera* Grown in Mexico. J Chem, 2015;860381. https://doi. org/10.1155/2015/860381

63. Fejér J, Kron I, Pellizzeri V, Pluchtová M, Eliašová A, Campone L, Gervasi T, Bartolomeo G, Cicero N, Babejová A, Konecná M, Sedlak V, Porácová J, Grulová D. First report on evaluation of basic nutritional and antioxidant properties of *Moringa oleifera* Lam. from Caribbean Island of Saint Lucia. Plants, 2019;8:537. https://doi.org/10.3390/plants8120537

64. Moyo B, Masika P, Hugo A, Muchenje V. Nutritional characterization of *Moringa (Moringa oleifera* Lam.) leaves, Afr J Biotechnol, 2011;10(2011):12925–12933. https://doi.org/10.3390/plants8120537

65. Castillo-López RI, León-Félix J, Angulo-Escalante MÁ, Gutiérrez-Dorado R, Muy-Rangel MD, Heredia JB. Nutritional and phenolic characterization of *Moringa oleifera* leaves grown in Sinaloa, México. Pak J Bot, 2017;49:161–168.

66. Fokwen VF, Tsafack HD, Touko BAH, Djopnang D, Afeanyi TA, Kong AT, Djikeng FT, Womeni HM. Nutrients composition, phenolic content and antioxidant activity of green and yellow *Moringa (Moringa oleifera)* leaves. J Food Stab, 2019;1:46–56.

67. Yameogo CW, Bengaly MD, Savadogo A, Nikiema PA, Traore SA. Determination of chemical composition and nutritional values of *Moringa oleifera* leaves. Pak J Nut, 2011;10(3):264–268.

68. Witt KA. The nutrient content of *Moringa oleifera* leaves. Messiah College Department of Nutrition and Dietetics, 2013.

69. Isitua CC, Lozano MJS, Jaramillo C, Dutan F. Phytochemical and nutritional properties of dried leaf powder of *Moringa oleifera* Lam. from Machala el oro province of Ecuador. Asian J Pl Sci Res, 2015;5:8–16.

70. Gopalakrishnan L, Doriya K, Kumar DS. *Moringa oleifera*: A review on nutritive importance and its medicinal application. Food Sci Hum Well, 2016;5(2):49–56. https://doi. org/10.1016/j.fshw.2016.04.001

71. Uphadek B, Shinkar DM, Patil PB, Saudagar RB. *Moringa oleifera* as a pharmaceutical excipient. Int J Curr Pharm Res, 2018;10(2):13–16. https://doi.org/10.22159/ ijcpr.2018v10i2.25883

72. Vergara-Jimenez M, Almatrafi MM, Fernandez M. Bioactive components in *Moringa oleifera* leaves protect against chronic disease. Antioxidants, 2017;6(9):1–13. https://doi. org/10.3390/antiox6040091

73. Ayoade ET, Akinyemi OA, Oyelere FS. Phytochemical profile of different morphological organs of *Moringa oleifera* plant. The J Phytopharmacol, 2019;8(6):295–298. https://doi. org/10.31254/phyto.2019.8605

74. Sudha R, Philip XC, Suriyakumari KVP. Phytochemical constituents of leaves of *Moringa oleifera* grown in Cuddalore District, Tamil Nadu, India. SBV J Basic, Clin Appl Health Sci, 2020;3(4):164–167. https://doi.org/10.5005/jp-journals-10082-02270

75. Koruthu DP, Manivarnan NK, Gopinath A, Abraham R. Antibacterial evaluation, reducing power assay and phytochemical screening of *Moringa oleifera* leaf extracts: effect of solvent polarity. Int J Pharm Sci Res, 2011;2(11):2991–2995. http://dx.doi.org/10.13040/ IJPSR.0975-8232.2(11).2991-95

76. Udikala M, Verma Y, Sushma, Lal S. Phytonutrient and pharmacological significance of *Moringa oleifera*. Int J Life Sci Scient Res, 2017;3(5):1387–91.

77. Jain P, Jain N, Patil UK. Phytochemical and pharmacological profile of *Moringa oleifera* Lam. Int J Pharm Sci Res, 2020; 11(12):5968–5973. http://dx.doi.org/10.13040/ IJPSR.0975-8232.11(12).5968-73

78. Mishra G, Singh P, Verma R, Kumar S, Srivatsav S, Jha KK, Khosa RL. Traditional uses, phytochemistry and pharmacological properties of *Moringa oleifera* plant: An overview. Der Pharm Lett, 2011;3(2):141–164.

79. Reminus O, Cornelius W. Phytochemical analysis of *Moringa oleifera* (leaves and flowers) and the functional group. Global Scient J, 2019;7(6):41–51.

80. Djenidi H, Khennouf S, Bouaziz A. Antioxidant activity and phenolic content of commonly consumed fruits and vegetables in Algeria. Prog Nut, 2020;22(1):224–235. http:// dx.doi.org/10.23751/pn.v22i1.7701

81. Abdulkadir A R, Zawawi DD, Md. Sarwar Jahan. DPPH antioxidant activity, total phenolic and total flavonoid content of different part of Drumstic tree (*Moringa oleifera* Lam.). J Chem Pharm Res, 2015;7(4):1423–1428.

82. Sultana B, Anwar F. Flavonols (kaempeferol, quercetin, myricetin) contents of selected fruits, vegetables and medicinal plants. Food Chem, 2008;108(3):879–884. http://dx.doi. org/10.1016/j.foodchem.2007.11.053

83. Gunathilake KDPP, Ranaweera KKDS. Antioxidative properties of 34 green leafy vegetables. J Funct Foods, 2016;26:176–186. https://doi.org/10.1016/j.jff.2016.07.015

84. Stohs SJ, Hartman MJ. Review of the safety and efficacy of *Moringa oleifera*. Phytother Res, 2015;29(6):796–804. https://doi.org/10.1002/ptr.5325

85. Lin H. Comparative analysis of chemical constituents of *Moringa oleifera* leaves from China and India by ultra-performance liquid chromatography coupled with quadrupole-time-of-flight mass spectrometry. Molecules, 2019;24(942):1–25. https://doi.org/10.3390/molecules24050942

86. Lin M, Zhang J, Chen X. Bioactive flavonoids in *Moringa oleifera* and their health-promoting properties. J Fun Foods, 2018;47:469–79. https://doi.org/10.1016/j.jff.2018.06.011

87. Masood S, Rehman A, Ihsan MA, Shahzad K, Sabir M, Alam S, Ahmed W, Shah ZH, Alghabari F, Mehmood A, Chung G. 2023. Antioxidant potential and α-glucosidase inhibitory activity of onion (*Allium cepa* L.) peel and bulb extracts. Brazil J Biol, 2023;83(e247168): 1–9. https://doi.org/10.1590/1519-6984.247168

88. Hussain A. A phylogenetic perspective of antiviral species of the genus *Artemisia* (Asteraceae-Anthemideae): A proposal of anti SARS-CoV-2 (COVID-19) candidate taxa. J Herb Med, 2022;36(2022):100601. https://doi.org/10.1016/j.hermed.2022.100601

89. Zhu Y, Yin Q, Yang Y. Comprehensive investigation of *Moringa oleifera* from different regions by simultaneous determination of 11 polyphenols using UPLC-ESI-MS/MS. Molecules, 2020;25:676. https://doi.org/10.3390/molecules25030676

90. Marcel A, Hubert M, Bienvenu MJ, Pascal O. 2016. Physico-chemical characteristics and biochemical potential of *Moringa oleifera* Lam. (Moringaceae). Der Pharm Lett, 2016;8(18):43–47.

91. Rubio-Sanz L, Dorca-Fornell C, Ornos MF, Navarro-León E, Jaizme-Vega MC. Phytochemical characterization of *Moringa oleifera* leaves. Herb Polon, 2021;67(3):19–26. https:// doi.org/10.2478/hepo-2021-0019

92. Asogwa IS, Ani JC. Effect of *Moringa oleifera* leaf powder inclusion on the phytochemical and antioxidant activity of *Akamu*. Agro-Sci, 2017;16(2):23–30. https://doi.org/10.4314/as.v16i2.4

93. Hussain A. A preliminary up-to-date review on Pakistani medicinal plants with potential antioxidant activity. RADS J Biol Res Appl Sci, 2020;11(1):61–88. https://doi.org/10.37962/jbas.v11i1.275

94. Dastmalchi K, Dorman HJD, Kosar M, Hiltunen R. Chemical composition and in vitro antioxidant evaluation of a water soluble Moldavian balm (*Dracocephalum moldavica* L.) extract. Leben Wissens Technol, 2007;40:239–-248. https://doi.org/10.1016/j.lwt.2005.09.019

95. Saeed MK, Zahra N, Saeed A, Abidi SHI, Syed Q. *Syzygium Cumini* L. seed, a potent source of fiber, protein and natural antioxidants. LGU J Life Sci, 2022c;6(3):252-256 https://doi.org/10.54692/lgujls.2022.0603227

96. Alallan L. Study of the chemicals, phenols, flavonoids, and antioxidants content of the Syrian *Arum hygrophilum* Boiss plant. Int J Herb Med, 2021;9(6):62–66.

97. Karim I, Khalid M, Mughal AA. Comparative study of antioxidative properties of Jhelum valley fruits. Pure Appl Biol, 2022;11(3):861–870. http://dx.doi.org/10.19045/ bspab.2022.110088

98. Al-Taweel SK, Al-Anbari IH. *Moringa olifera*: a review on the phytochemical screening, proximate analysis, medicinal, nutritional, and plant biostimulants values of its leaves, pods, seeds and roots. Pl Arch, 2019;19(2):1612–1622.

99. Ali A, Garg P, Goyal R, Kaur G, Li X, Negi P, Valis M, Kuca K, Kulshrestha S. A novel herbal hydrogel formulation of *Moringa oleifera* for wound healing. Plants, 2021;10(1):25. http://dx.doi.org/10.3390/plants10010025

100. Kinyi HW, Tirwomwe M, Ninsiima HI, Miruk CO. Effect of cooking method on vitamin C loses and antioxidant activity of indigenous green leafy vegetables consumed in Western Uganda. Int J Food Sci, 2022;1-7. http://dx.doi.org/10.1155/2022/2088034

101. Kumar V, Pandey N, Mohan N, Singh RP. Antibacterial and antioxidant activity of different extract of *Moringa oleifera* leaves. Int J Pharm Sci Rev Res, 2012b;12:89–94.

102. Almaghrabi AM, Kadasa OA, Mohamed A.A. Antioxidant and antimicrobial potential of *Moringa oleifera* extract against food pathogens. Biosci Biotechnol Res Comm, 2021;14(3):1098–1104.

103. Landázuri AC, Gualle A, Castañeda V, Morales E, Caicedo A, Orejuela-Escobar LM. *Moringa oleifera* Lam. leaf powder antioxidant activity and cytotoxicity in human primary fibroblasts. Nat Prod Res, 2021;35(24):6194–6199. http://dx.doi.org/10.1080/14786419.20 20.1837804

104. Khor KZ, Lim V, Moses EJ, Abdul Samad N. The *in vitro* and *in vivo* anticancer properties of *Moringa oleifera*. Evid Based Compl Alt Med, 2018;10(7):1243–1252. https://doi.org/10.1155/2018/1071243

105. Saleem A, Saleem M, Akhtar MF. Antioxidant, anti-inflammatory and antiarthritic potential of *Moringa oleifera* Lam: An ethnomedicinal plant of Moringaceae family. South Afr J Bot, 2020;128:246–256. https://doi.org/10.1016/j.sajb.2019.11.023

106. Hussain A, Sajid M, Rasheed H, Hassan M, Khan MA, Bokhari SAI. Phytochemistry and antibacterial efficacy of Northeastern Pakistani *Artemisia rutifolia* Stephan ex Spreng. extracts against some clinical and phyto-pathogenic bacterial strains. Acta Pharm Sci, 2022;60;247–271. https://doi.org/10.23893/1307- 2080.APS.6017

107. Elazzazy AM, Almaghrabi OA, Kadasa NMS, Mohamed AA. Antioxidant and antimicrobial potential of *Moringa oleifera* extract against food pathogens. Biosci Biotechnol Res Commun, 2021; 14(3):1–12. http://dx.doi.org/10.21786/bbrc/14.3.29

108. Flieger J, Flieger M. The [DPPH•/DPPH-H]-HPLC-DAD method on tracking the antioxidant activity of pure antioxidants and goutweed (*Aegopodium podagraria* L.) hydroalcoholic extracts. Molecules, 2020;25(24):6005. https://doi.org/10.3390/molecules25246005

109. Yadav N, Pal A, Sihag S, Nagesh CR. Antioxidant activity profiling of acetonic extract of jamun (*Syzygium cumini* L.) seeds in different *in-vitro* models. Open Food Sci J, 2020;12:3–8. https://doi.org/10.2174/1874256402012010003

110. Wajiha, Qureshi NA. *In vitro* anticoccidial, antioxidant activities and biochemical screening of methanolic and aqueous leaves extracts of selected plants. Pak Vet J, 2021;41(1):57– 63. https://doi.org/10.29261/pakvetj/2020.071

111. Jomi JA, Abarna S, Sathishkumar T, Baskar R, Muthukumaran P. Extraction, evaluation, and antioxidant activity of total phenol from callus of *Abutilon indicum* (L.) Sweet. Lett Appl NanoBiosci, 2022;11(3):3652–3660. https://doi.org/10.33263/lianbs113.36523660